

## **CELL LINE CHARACTERIZATION TESTING INFORMATION**

| Assay   | What it Detects   | What it Doesn't Detect  | TAT <sup>1</sup>                     | When to Use  |
|---|---|---|--------------------------------------|--|
| G-Banded Karyotyping                                | Microscopic genomic abnormalities (>5-10 Mb) Inversions Duplications/deletions Balanced and unbalanced translocations Aneuploidies >10% mosaicism (e.g. cultures where 1 of 10 cells are trisomy 12)  | Submicroscopic genomic abnormalities (<5 Mb) <~10% culture mosaicism (e.g. cultures where 1 of 10 cells is trisomy 12)  | 7-10 days<br>(4-6 days<br>expedited) | As a baseline genomic screen  At derivation of cell lines  At the start of experimental protocols  To assess and monitor genomic stability (e.g. every 5-10 passages of cell culture)  At conclusion of experiments (prior to publication)  For cell line banking  When publication-quality karyotypes are needed  |
| Fluorescence <i>In Situ</i> Hybridization<br>(FISH) | Genomic sequence of interest Duplications or deletions >100 Kb >2% mosaicism (e.g. cultures where >2 of 100 cells are trisomy 12) Chromosomal location of genomic gains Chromosome fusions (breakaparts)  | Changes in regions other than the probe-<br>specific sequence   | 10-15 days                           | To confirm findings and refine breakpoints detected by g-banded karyotyping To confirm findings and localize genomic gains detected by SNP microarray As a screen for microdeletions/duplications of known targets   |
| Short Tandem Repeat Analysis (STR)                  | STR polymorphisms for 15 loci plus amelogenin<br>(Promega® PowerPlex® 16 HS)     Probability of matching identity to an existing<br>STR profile   | STR polymorphisms in areas other than those represented in Promega® PowerPlex® 16 HS  | 10-20 days                           | <ul> <li>To monitor identity of a cell line</li> <li>To confirm relationship of iPS cells to their parent line</li> <li>To establish a STR profile of a newly derived or reprogrammed cell line</li> <li>To rule out culture cross-contamination</li> </ul>  |
| Mycoplasma Detection by PCR                         | 96 species of mycoplasma contamination from stem cell cultures.     Sensitivity (5-100 CFU/ml)  | This system does not allow for the amplification of DNA originating from other sources, such as bacteria.   | 5-7 days                             | To monitor the health of your cell line To monitor for contamination in shared lab spaces To assure that mycoplasma is not interfering with your experiments To rule out mycoplasma as the culprit of chromosomal aberrations  |
| Single nucleotide polymorphism (SNP)<br>Microarray  | Submicroscopic genomic abnormalities (>5-10 Mb) Genomic gains and losses (>50 Kb) Copy number variants Duplications/deletions Unbalanced translocations Aneuploidies Copy neutral Loss of Heterozygosity (LOH) / Absence of Heterozygosity (AOH) (>5 Mb)  >~10% mosaicism (e.g. cultures where >1 of 10 cells are trisomy 12) | Balanced translocations Robertsonian Balanced insertions Inversions <~10% culture mosaicism (e.g. cultures where 1 of 10 cells is trisomy 12) Chromosomal position of genomic gains | 14-21 days                           | As a baseline genomic screen     To detect submicroscopic (<5 Mb) abnormalities     To identify amplified or deleted genes of interest     To assess and monitor genomic stability     (e.g. every 5-10 passages of cell culture)  In conjunction with G-banded karyotyping     To define unbalanced translocation breakpoints  For research of genomic copy number change     To identify structural variation within     populations or disease cohorts     To develop a cell line copy number variant profile |
| Spectral Karyotyping (SKY)                          | Microscopic genomic abnormalities (>5-10 Mb)     Balanced and unbalanced translocations     Aneuploidies  | <ul> <li>Submicroscopic genomic abnormalities (&lt;5 Mb)</li> <li>Inversions</li> <li>Duplications/deletions</li> </ul>   | 14-21 days                           | As an adjunct to g-banded karyotyping     To define complex rearrangements     To identify marker chromosomes     When publication-quality spectral karyotypes are needed  |

<sup>&</sup>lt;sup>1</sup>Turn-around-times (TAT) provided are based on provision of sufficient mitotically active hES or iPS cultures grown in Matrigel/TeSR or MEF/hES media conditions. Turn-around-times may be longer for cGMP compliant testing.