




# Certificate of Analysis

## Thaw and Culture Details

Cell Line Name	<b>CVCL_C7V1</b>	
WiCell Lot Number	<b>WB68881</b>	
Provider/Client	Albert Einstein College of Medicine – Dr. Frank Soldner	
Banked By	WiCell	
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 2 well of a 6 well plate using iSCORE 01 medium and MEF. WiCell recommends thawing using ROCK Inhibitor for best results.	
Protocol	WiCell Feeder Based (MEF) Protocol 01 for Culture of MJFF iSCORE Lines	
Culture Platform Prior to Freeze	Medium: iSCORE 01 medium	Matrix: MEF
Passage Number	p42 Cells were cultured for 41 passages prior to freeze and post colony selection. Plated cells at thaw should be labeled passage 42.	
Date Vialled	14-May-2025	
Vial Label	CVCL_C7V1 p42 WB68881  Store at -135C or colder Made in United States Research Use Only 	
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.	



# Certificate of Analysis

## Results

Test Description	Test Provider	Test Method	Test Specification	Result
Karyotype	WiCell	G-T-L Banding performed on 20 metaphase cells	Expected karyotype	See Report
	<b>Results:</b> 46,XX <b>Interpretation:</b> This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.			
Post-Thaw Viable Cell Recovery	WiCell	Thaw using specified Thaw & Culture Recommendations	≥ 15 Undifferentiated Colonies prior to passage, ≤ 30% Differentiation prior to passage, and recoverable attachment after passage	Pass
Identity by STR	WiCell	PowerPlex 16 HS System by Promega™	Consistent with STR profile of deposited cell line	See Report
Mycoplasma	WiCell	PCR	Amplification of mycoplasma specific DNA detected with negative result	Pass
Sterility	Steris	Native Product Direct Transfer using FTM and TSB (ST/07)	Negative for growth following 14 days of culture	Pass

Approval Date	WiCell Quality Assurance Approval
20-November-2025	<div>11/20/2025</div> <div>X HEB</div> <div>HEB WiCell Quality Assurance Signed by: Bruner, Holley</div>

The material provided under this certificate has been subjected to the tests specified and the results and data described herein are accurate based on WiCell's reasonable knowledge and belief. Appropriate Biosafety Level practices and universal precautions should always be used with this material. For clarity, the foregoing is governed solely by WiCell's Terms and Conditions of Service, which can be found at <http://www.wicell.org/privacyandterms>.

**Date Reported:** June 15, 2025

**Cell Line:** CVCL\_C7V1-WB68881

**Submitted Passage #:** 43

**Date of Sample:** 6/6/2025

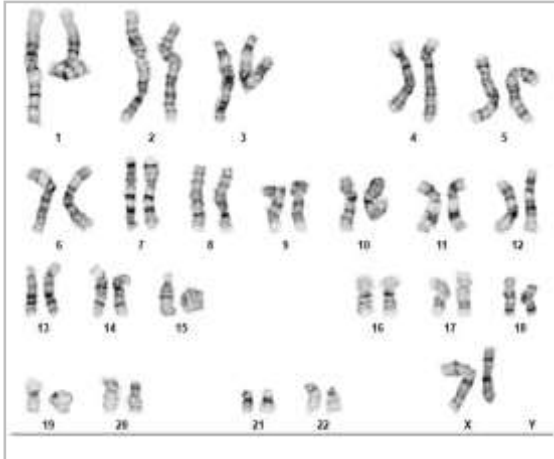
**Specimen:** Human ESC

**Results:** 46,XX

**Cell Line Sex:** Female

**Reason for Testing:** LOT\_RELEASE

**Investigator:** WiCell Stem Cell Bank, WiCell



**Cell:** 12

**Slide:** G01

**Slide Type:** Karyotype

**Total Counted:** 16

**Total Analyzed:** 8

**Total Karyogrammed:** 4

**Band Resolution:** 400 - 475

## Interpretation:

**This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.**

**This is a limited analysis, based on sixteen cells examined. Standard analysis requires examination of twenty cells. All analyzable metaphase cells were evaluated.**

**Completed by:** Pam Mill

**Reviewed and Interpreted by:** Justin Schleede, PhD, FACMG

For internal use only

**Date:** \_\_\_\_\_ **Sent By:** \_\_\_\_\_ **Sent To:** \_\_\_\_\_ **QC Review By:** \_\_\_\_\_

*Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".*

*This assay was conducted solely for listed investigator/institution. The results of this assay are for research use only. Unless otherwise mutually agreed in writing, the services provided to you hereunder by WiCell Research Institute, Inc. ("WiCell") are governed solely by WiCell's Terms and Conditions of Service, found at [www.wicell.org/privacyandterms](http://www.wicell.org/privacyandterms). Any terms you may attach to a purchase order or other document that are inconsistent, add to, or conflict with WiCell's Terms and Conditions of Service are null and void and of no legal force or effect.*



# Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell  
Sample Receipt Date: 12Jun25, 10Jun25, 06Jun25, 02Jun25  
STR Amplification Date: 12Jun25, 13Jun25

Form SOP-89.01  
Version 15.0

Sample Name	CVCL_C7V7- WB68815 p40	PACSLi003-A- DB68806 p9	CVCL_C7V8- WB68846 p40	CVCL_C7V5- WB68880 p38	BCHi013-A-1- DB68699 p20	CVCL_C7V1- WB68881 p43
WiCell CTR No. <sup>1</sup>	Identifying information has been redacted to protect donor confidentiality. If more information is required, please contact <a href="mailto:info@wicell.org">info@wicell.org</a>					
FGA						
TPOX						
D8S1179						
vWA						
Amelogenin						
Penta_D						
CSF1PO						
D16S539						
D7S820						
D13S317						
D5S818						
Penta_E						
D18S51						
D21S11						
TH01						
D3S1358						
Allelic Polymorphisms	28	28	28	28	25	28
Matches <sup>2</sup>	See Results		See Results	See Results		See Results
Comments						

<sup>1</sup> CTR No.: Characterization Test Request Number; also known as a laboratory accessioning number.

<sup>2</sup> The STR profile of the sample(s) listed are a 100% match for the given sample unless otherwise specified.



# Short Tandem Repeat

Form SOP-89.01  
Version 15.0

Requestor: WiCell Stem Cell Bank, WiCell  
Sample Receipt Date: 12Jun25, 10Jun25, 06Jun25, 02Jun25  
STR Amplification Date: 12Jun25, 13Jun25

**Assay Description:** Short Tandem Repeat (STR) analysis is performed using the PowerPlex® 16 HS System by Promega™. Results are reported as 13 CODIS STR markers, Amelogenin for sex determination and two low-stutter, highly discriminating pentanucleotide STR markers.

**Results:** The genotypic profiles comprise a range of 25-28 allelic polymorphisms across the 15 STR loci analyzed. Samples 107971, 107881, 107916 and 107917 are a 100% match to each other and to 105684, 105942, 107124, 107305, 107471, 107640 and additional profiles. Additional matches can be provided upon request.

**Interpretation:** The concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggest that the cells submitted correspond to the cell lines as named and were not contaminated with any other human cells or a significant amount of mouse feeder layer cells.

**Sensitivity:** Sensitivity limits for detection of STR polymorphisms unique to either this or other human cell lines is ~2-4%.

6/17/2025	6/18/2025	6/19/2025
<b>X</b> Amber Kuhn	<b>X</b> Anna Lisa Larson	<b>X</b> Hunter Hefti
Tech #1 Characterization Signed by: Kuhn, Amber	Tech #2 Characterization Signed by: Larson, Anna Lisa	QA Review Quality Assurance Signed by: Hefti, Hunter

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# Mycoplasma Assay Report

PCR-based assay performed by WiCell  
WiCell Stem Cell Bank, WiCell  
17Jun25

Form SOP-83.01  
Version 7.0

Sample Name	Result	Interpretation
CVCL_C7V1-WB68881 p43 (107881)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 995 09Jun25 NM (107898)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 995 09Jun25 JG (107899)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 995 09Jun25 KC (107900)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 815 09Jun25 KC (107901)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 841 09Jun25 AP (107902)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 994 09Jun25 KC (107903)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 994 09Jun25 JG (107904)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Incubator 994 09Jun25 NM (107905)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
CVCL_C7VI-DB68458 p31 (107937)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Positive (+) Control	Positive	
Negative (-) Control	Negative	

## Assay Description

Sample is tested for presence of mycoplasma using EZ-PCR™ Mycoplasma Detection Kit (Sartorius).

6/17/2025  <b>X</b> Steph Dos Santos Tech #1 Characterization Signed by: Dos Santos, Stephany	6/17/2025  <b>X</b> Nina Montgomery Tech #2 Characterization Signed by: Montgomery, Nina	6/18/2025  <b>X</b> Hunter Hefti QA Review Quality Assurance Signed by: Hefti, Hunter
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A gel image is available upon request.

# Native Product Sterility Report



WiCell Research Institute  
504 S Rosa Road, Rm 101  
Madison, WI 53719

SAMPLE #: 25060863  
DATE RECEIVED: 26-Jun-25  
TEST INITIATED: 27-Jun-25  
TEST COMPLETED: 11-Jul-25

SAMPLE NAME / DESCRIPTION:

BCHi018-A-9-DB68713  
BCHi019-A-10-DB68716  
BCHi019-A-11-DB68717  
BCHi019-A-9-DB68715  
BCHi020-A-5-DB68718  
BCHi020-A-8-DB68719  
BCHi020-A-9-DB68720  
EIFIIIi001-A-DB68807  
EIFIIIi002-A-DB68808  
PACSIi002-A-DB68805  
PACSIi003-A-DB68806  
CVCL\_C7V1-WB68881  
CVCL\_C7V2-WB68887  
CVCL\_C7V3-WB68899  
CVCL\_C7V4-WB68879  
CVCL\_C7V5-WB68880  
CVCL\_C7V6-WB68883  
CVCL\_C7V7-WB68815  
CVCL\_C7V8-WB68846  
CVCL\_C7VS-WB68804

UNIQUE IDENTIFIER: N/A

## TEST RESULTS:

# Tested	# Positives (Growth)	- Control
20	0	2 Negatives

## TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
20	TSB	40	20-25	14
20	FTG	40	30-35	14

# Native Product Sterility Report



REFERENCE: Processed according to LAB-003: Sterility Test Procedure

PD #: 000053

TEST METHODOLOGY: USP - Direct Transfer

COMMENTS: SAMPLE#: 25060863

Upon receipt the samples listed below contained 0.5 mL of fluid, 0.25 mL was tested in each media.

EIFIIIi001-A-DB68807

EIFIIIi002-A-DB68808

PACSIli002-A-DB68805

PACSIli003-A-DB68806

AUTHORIZED BY

A handwritten signature in blue ink, appearing to be "A. R.", written over a horizontal line.

DATE

16 JUL 2025

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests. Results applied to samples as received.