



Certificate of Analysis

Thaw and Culture Details

Cell Line Name	CVCL_C7VI	
WiCell Lot Number	WB69048	
Provider/Client	Albert Einstein College of Medicine – Dr. Frank Soldner	
Banked By	WiCell	
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 1 well of a 6 well plate using iSCORE 01 medium and MEF. WiCell recommends thawing using ROCK Inhibitor for best results.	
Protocol	WiCell Feeder Based (MEF) Protocol 01 for Culture of MJFF iSCORE Lines	
Culture Platform Prior to Freeze	Medium: iSCORE 01 medium	Matrix: MEF
Passage Number	p42 Colony selection occurred at passage 24. Cells were cultured for 17 passages prior to freeze and post colony selection. Plated cells at thaw should be labeled passage 42.	
Date Vialed	06-August-2025	
Vial Label	CVCL_C7VI p42 WB69048 Store at -135C or colder Made in United States Research Use Only	
WiCell®		
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.	

The material provided under this certificate has been subjected to the tests specified and the results and data described herein are accurate based on WiCell's reasonable knowledge and belief. Appropriate Biosafety Level practices and universal precautions should always be used with this material. For clarity, the foregoing is governed solely by WiCell's Terms and Conditions of Service, which can be found at <http://www.wicell.org/privacyandterms>.



Certificate of Analysis

Results

Test Description	Test Provider	Test Method	Test Specification	Result
Karyotype	WiCell	G-T-L Banding performed on 20 metaphase cells	Expected karyotype	See Report
Results: 46,XX Interpretation: This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.				
Post-Thaw Viable Cell Recovery	WiCell	Thaw using specified Thaw & Culture Recommendations	≥ 15 Undifferentiated Colonies prior to passage, ≤ 30% Differentiation prior to passage, and recoverable attachment after passage	Pass
Identity by STR	WiCell	PowerPlex 16 HS System by Promega™	Defines STR profile of deposited cell line	See Report
Mycoplasma	WiCell	PCR	Amplification of mycoplasma specific DNA detected with negative result	Pass
Sterility	Steris	Native Product Direct Transfer using FTM and TSB (ST/07)	Negative for growth following 14 days of culture	Pass

Approval Date	WiCell Quality Assurance Approval
02-January-2026	<p>1/2/2026</p> <p>X Jenna Gay</p> <p>Jenna Gay WiCell Quality Assurance Signed by: Gay, Jenna</p>

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Date Reported: October 08, 2025

Cell Line Sex: Female

Cell Line: CVCL_C7VI-WB69048

Reason for Testing: LOT_RELEASE

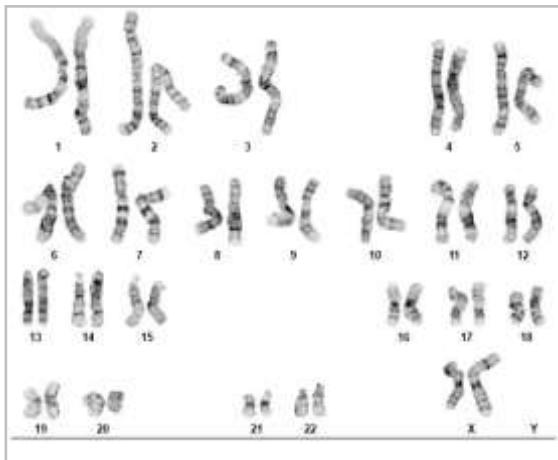
Submitted Passage #: 46

Date of Sample: 10/6/2025

Investigator: WiCell Stem Cell Bank, WiCell

Specimen: Human Modified ESC

Results: 46,XX



Cell: 36

Slide: G03

Slide Type: Karyotype

Total Counted: 20

Total Analyzed: 8

Total Karyogrammed: 4

Band Resolution: 400 - 500

Interpretation:

This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.

Completed by: Davena Lira, CG(ASCP)

Reviewed and Interpreted by: Justin Schleede, PhD, FACMG

For internal use only

Date: _____ Sent By: _____ Sent To: _____ QC Review By: _____

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results of this assay are for research use only. Unless otherwise mutually agreed in writing, the services provided to you hereunder by WiCell Research Institute, Inc. ("WiCell") are governed solely by WiCell's Terms and Conditions of Service, found at www.wicell.org/privacyandterms. Any terms you may attach to a purchase order or other document that are inconsistent, add to, or conflict with WiCell's Terms and Conditions of Service are null and void and of no legal force or effect.



Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell

Form SOP-89.01

Version 15.0

Sample Receipt Date: 04Oct25, 06Oct25, 07Oct25, 08Oct25, 09Oct25

STR Amplification Date: 14Oct25

Sample Name	SUSL-046_C2-DB68947 p12	CVCL_C7VI-WB69048 p46	EIFIIIi002-A-WB69082 p11	SUSL-025_C2-DB68955 p16	SUSL-046_C9-DB68948 p11	SUSL-047_C5-DB68950 p12
WiCell CTR No. ¹						
FGA						
TPOX						
D8S1179						
vWA						
Amelogenin						
Penta_D						
CSF1PO						
D16S539						
D7S820						
D13S317						
D5S818						
Penta_E						
D18S51						
D21S11						
TH01						
D3S1358						
Allelic Polymorphisms	27	28	27	26	27	26
Matches ²	109464	See Results	108779		109398	109466 (96.67%)
Comments						

¹ CTR No.: Characterization Test Request Number; also known as a laboratory accessioning number.

² The STR profile of the sample(s) listed are a 100% match for the given sample unless otherwise specified.



Short Tandem Repeat

Form SOP-89.01

Version 15.0

Requestor: WiCell Stem Cell Bank, WiCell

Sample Receipt Date: 04Oct25, 06Oct25, 07Oct25, 08Oct25, 09Oct25

STR Amplification Date: 14Oct25

Sample Name	SUSL-047_C8-DB68951 p12
WiCell CTR No. ¹	
FGA	
TPOX	
D8S1179	
vWA	
Amelogenin	
Penta_D	
CSF1PO	
D16S539	
D7S820	
D13S317	
D5S818	
Penta_E	
D18S51	
D21S11	
TH01	
D3S1358	
Allelic Polymorphisms	
Matches ²	109465 (96.67%)
Comments	

¹ CTR No.: Characterization Test Request Number; also known as a laboratory accessioning number.

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Short Tandem Repeat

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Sample Receipt Date: 04Oct25, 06Oct25, 07Oct25, 08Oct25, 09Oct25

STR Amplification Date: 14Oct25

Assay Description: Short Tandem Repeat (STR) analysis is performed using the PowerPlex® 16 HS System by Promega™. Results are reported as 13 CODIS STR markers, Amelogenin for sex determination and two low-stutter, highly discriminating pentanucleotide STR markers.

Results: The genotypic profiles comprise a range of 26 to 28 allelic polymorphisms across the 15 STR loci analyzed. Samples 109407 is a 100% match to 109139, 109380, 109332, 109197, 109232, 109158, 108776, 108996, 108953, 108863, and additional profiles. Additional matches can be provided upon request.

Interpretation: The concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggest that the cells submitted correspond to the cell lines as named and were not contaminated with any other human cells or a significant amount of mouse feeder layer cells.

Sensitivity: Sensitivity limits for detection of STR polymorphisms unique to either this or other human cell lines is ~2-4%.

10/20/2025	10/20/2025	10/22/2025
X Steph Dos Santos	X Michael Mussar	X Hunter Hefti
Tech #1 Characterization Signed by: Dos Santos, Stephany	Tech #2 Characterization Signed by: Mussar, Michael	QA Review Quality Assurance Signed by: Hefti, Hunter

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Mycoplasma Assay Report

PCR-based assay performed by WiCell
WiCell Stem Cell Bank, WiCell

13Oct25

Form SOP-83.01
Version 7.0

Sample Name	Result	Interpretation
EIFIIIi002-A-WB69082 p11 (109415)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
EIFIIIi001-A-WB69083 p11 (109414)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
CVCL_C7VI-WB69048 p46 (109407)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Positive (+) Control	Positive	
Negative (-) Control	Negative	

Assay Description

Sample is tested for presence of mycoplasma using EZ-PCR™ Mycoplasma Detection Kit (Sartorius).

10/13/2025 X Kami Clerkin <hr/> Tech #1 Characterization Signed by: Kami Clerkin	10/13/2025 X Steph Dos Santos <hr/> Tech #2 Characterization Signed by: Dos Santos, Stephany	10/14/2025 X Hunter Hefti <hr/> QA Review Quality Assurance Signed by: Hefti, Hunter
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A gel image is available upon request.

Native Product Sterility Report



WiCell Research Institute
504 S Rosa Road, Rm 101
Madison, WI 53719

CORRECTED REPORT

SAMPLE #: 25080540
DATE RECEIVED: 14-Aug-25
TEST INITIATED: 15-Aug-25
TEST COMPLETED: 29-Aug-25

SAMPLE NAME / DESCRIPTION: CVCL_C7VF-WB69010
CVCL_C7VD-WB69009
BCHi014-A-9-WB69007
CVCL_C7VL-WB68996
CVCL_C7VG-WB68944
CVCL_C7VJ-WB69026
CVCL_C7UV-WB69028
BCHi014-A-10-WB69032
CVCL_C7UX-WB69034
CVCL_C7UW-WB69046
CVCL_C7VI-WB69048
WA14-WB69050
EIFIIIi001-A-DB68807
EIFIIIi002-A-DB68808
PACSIIIi002-A-DB68805
PACSIIIi003-A-DB68806

UNIQUE IDENTIFIER: N/A

TEST RESULTS:

# Tested	# Positives (Growth)	- Control
16	0	2 Negatives

TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
16	TSB	40	20-25	14
16	FTG	40	30-35	14

REFERENCE:

Processed according to LAB-003: Sterility Test Procedure

PD #:

000053

TEST METHODOLOGY:

USP - Direct Transfer

Native Product Sterility Report



COMMENTS: Report revised due to updated Sample Name/Description information.

CORRECTED REPORT

AUTHORIZED BY

A handwritten signature in blue ink that reads "Lee Vang".

DATE 09 Oct 25

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests. Results applied to samples as received.