Certificate of Analysis

Thaw and Culture Details

Cell Line Name	**CVCL_C7V7**	
WiCell Lot Number	**WB68815**	
Provider/Client	Albert Einstein College of Medicine – Dr. Frank Soldner	
Banked By	WiCell	
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 1 well of a 6 well plate using iSCORE 01 medium and MEF. WiCell recommends thawing using ROCK Inhibitor for best results.	
Protocol	WiCell Feeder Based (MEF) Protocol 01 for Culture of MJFF iSCORE Lines	
Culture Platform Prior to Freeze	Medium: iSCORE 01 medium	Matrix: MEF
Passage Number	p37 Colony selection occurred at passage 30. Cells were cultured for 6 passages prior to freeze and post colony selection. Plated cells at thaw should be labeled passage 37.	
Date Vialed	05-May-2025	
Vial Label	CVCL_C7V7 p37 WB68815 Store at -135C or colder Made in United States Research Use Only	
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.	

Certificate of Analysis

Results

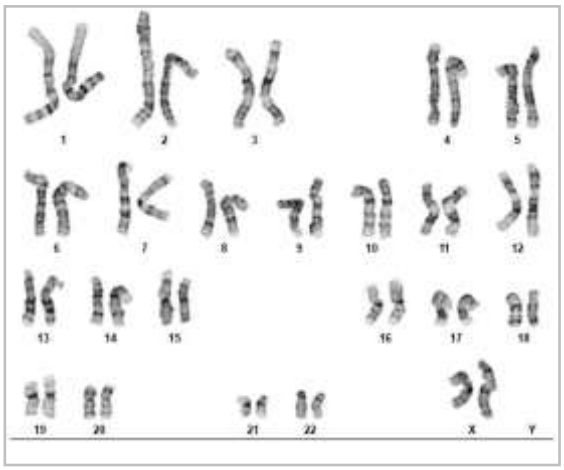
Test Description	Test Provider	Test Method	Test Specification	Result
Karyotype	WiCell	G-T-L Banding performed on 20 metaphase cells	Expected karyotype	See Report
	Results: 46,XX ***Interpretation:*** This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.			
Post-Thaw Viable Cell Recovery	WiCell	Thaw using specified Thaw & Culture Recommendations	≥ 15 Undifferentiated Colonies prior to passage, ≤ 30% Differentiation prior to passage, and recoverable attachment after passage	Pass
Identity by STR	WiCell	PowerPlex 16 HS System by Promega™	Defines STR profile of deposited cell line	See Report
Mycoplasma	WiCell	PCR	Amplification of mycoplasma specific DNA detected with negative result	Pass
Sterility	Steris	Native Product Direct Transfer using FTM and TSB (ST/07)	Negative for growth following 14 days of culture	Pass

Approval Date	WiCell Quality Assurance Approval
11-February-2026	2/11/2026 X HEB HEB WiCell Quality Assurance Signed by: Bruner, Haley

Chromosome Analysis Report: 108410

Date Reported: July 25, 2025
Cell Line: CVCL_C7V7-WB68815
Submitted Passage #: 40
Date of Sample: 7/15/2025
Specimen: Human ESC
Results: 46,XX

Cell Line Sex: Female
Reason for Testing: LOT_RELEASE
Investigator: WiCell Stem Cell Bank, WiCell



Cell: 5
Slide: G02
Slide Type: Karyotype

Total Counted: 20
Total Analyzed: 8
Total Karyogrammed: 4
Band Resolution: 425 - 475

Interpretation:
This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.

Completed by: Timm Gonzales, CG(ASCP)
Reviewed and Interpreted by: Justin Schleede, PhD, FACMG

For internal use only

Date:_____ **Sent By:**____ **Sent To:**_____ **QC Review By:** ____

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e.,mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell
Sample Receipt Date: 12Jun25, 10Jun25, 06Jun25, 02Jun25
STR Amplification Date: 12Jun25, 13Jun25

Form SOP-89.01
Version 15.0

Sample Name	CVCL_C7V7-WB68815 p40	PACSIIi003-A-DB68806 p9	CVCL_C7V8-WB68846 p40	CVCL_C7V5-WB68880 p38	BCHi013-A-1-DB68699 p20	CVCL_C7V1-WB68881 p43
WiCell CTR No.[1]	107971	107970	107917	107916	107779	107881
FGA	22, 23	[redacted]				
TPOX	8, 9					
D8S1179	11, 13					
vWA	14, 20					
Amelogenin	X, X					
Penta_D	9, 13					
CSF1PO	10, 12					
D16S539	11, 13					
D7S820	10, 11					
D13S317	11, 14					
D5S818	11, 12					
Penta_E	7, 18					
D18S51	16, 16					
D21S11	30, 32.2					
TH01	9.3, 9.3					
D3S1358	14, 17					
Allelic Polymorphisms	28	28	28	28	25	28
Matches[2]	See Results		See Results	See Results		See Results
Comments						

Identifying information has been redacted to protect donor confidentiality. If more information is required, please contact info@wicell.org

[1] CTR No.: Characterization Test Request Number; also known as a laboratory accessioning number.
[2] The STR profile of the sample(s) listed are a 100% match for the given sample unless otherwise specified.

Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell
Sample Receipt Date: 12Jun25, 10Jun25, 06Jun25, 02Jun25
STR Amplification Date: 12Jun25, 13Jun25

Assay Description: Short Tandem Repeat (STR) analysis is performed using the PowerPlex® 16 HS System by Promega™. Results are reported as 13 CODIS STR markers, Amelogenin for sex determination and two low-stutter, highly discriminating pentanucleotide STR markers.

Results: The genotypic profiles comprise a range of 25-28 allelic polymorphisms across the 15 STR loci analyzed. Samples 107971, 107881, 107916 and 107917 are a 100% match to each other and to 105684, 105942, 107124, 107305, 107471, 107640 and additional profiles. Additional matches can be provided upon request.

Interpretation: The concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggest that the cells submitted correspond to the cell lines as named and were not contaminated with any other human cells or a significant amount of mouse feeder layer cells.

Sensitivity: Sensitivity limits for detection of STR polymorphisms unique to either this or other human cell lines is ~2-4%.

6/17/2025 X Amber Kuhn Tech #1 Characterization Signed by: Kuhn, Amber	6/18/2025 X Anna Lisa Larson Tech #2 Characterization Signed by: Larson, Anna Lisa	6/19/2025 X Hunter Hefti QA Review Quality Assurance Signed by: Hefti, Hunter

Mycoplasma Assay Report

PCR-based assay performed by WiCell

WiCell Stem Cell Bank, WiCell

01Jul25

Form SOP-83.01
Version 7.0

Sample Name	Result	Interpretation
CVCL_C7VR-DB68427 p29 (108209)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
CVCL_C7VN-DB68439 p32 (108208)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
CVCL_C7V7-WB68815 p37 (108207)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
BCHi014-A-9-DB68701 p17 (108206)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Positive (+) Control	Positive	
Negative (-) Control	Negative	

Assay Description
Sample is tested for presence of mycoplasma using EZ-PCR™ Mycoplasma Detection Kit (Sartorius).

7/1/2025 X Steph Dos Santos Tech #1 Characterization Signed by: Dos Santos, Stephany	7/1/2025 X Nina Montgomery Tech #2 Characterization Signed by: Montgomery, Nina	7/2/2025 X Emily Weber QA Review Quality Assurance Signed by: Weber, Emily

A gel image is available upon request.

Native Product Sterility Report

STERIS

WiCell Research Institute
504 S Rosa Road, Rm 101
Madison, WI 53719

SAMPLE #: 25060863
DATE RECEIVED: 26-Jun-25
TEST INITIATED: 27-Jun-25
TEST COMPLETED: 11-Jul-25

SAMPLE NAME / DESCRIPTION:

BCHi018-A-9-DB68713
BCHi019-A-10-DB68716
BCHi019-A-11-DB68717
BCHi019-A-9-DB68715
BCHi020-A-5-DB68718
BCHi020-A-8-DB68719
BCHI020-A-9-DB68720
EIFIIIi001-A-DB68807
EIFIIIi002-A-DB68808
PACSIIi002-A-DB68805
PACSIIi003-A-DB68806
CVCL_C7V1-WB68881
CVCL_C7V2-WB68887
CVCL_C7V3-WB68899
CVCL_C7V4-WB68879
CVCL_C7V5-WB68880
CVCL_C7V6-WB68883
CVCL_C7V7-WB68815
CVCL_C7V8-WB68846
CVCL_C7VS-WB68804

UNIQUE IDENTIFIER: N/A

TEST RESULTS:

# Tested	# Positives (Growth)	- Control
20	0	2 Negatives

TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
20	TSB	40	20-25	14
20	FTG	40	30-35	14

Native Product Sterility Report

STERIS

REFERENCE: Processed according to LAB-003: Sterility Test Procedure

PD #: 000053

TEST METHODOLOGY: USP - Direct Transfer

COMMENTS: SAMPLE#: 25060863

Upon receipt the samples listed below contained 0.5 mL of fluid, 0.25 mL was tested in each media.
EIFIIIi001-A-DB68807
EIFIIIi002-A-DB68808
PACSIIi002-A-DB68805
PACSIIi003-A-DB68806

AUTHORIZED BY _____ DATE 16JUL2025

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests.Results applied to samples as received.