Thaw and Culture Details

Cell Line Name	**iPS DF19-9-7T-PCBC**
WiCell Lot Number	**DB36826**
Provider	University of Wisconsin – Laboratory of Dr. James Thomson
Banked By	National Heart, Lung and Blood Institute Progenitor Cell Biology Consortium (NHLBI PCBC)
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 1 well of a 6 well plate. The Provider recommends only dispase passaging.
Culture Platform	Feeder Independent
	Medium: mTeSR™1
	Matrix: Matrigel®
Protocol	WiCell Feeder Independent mTeSR™1 Protocol
Passage Number	p33+9 These cells were cultured for 32 passages prior to freeze, at least 8 of them in mTeSR™1/Matrigel®. Therefore, plated cells at thaw should be labeled passage 33.
Date Vialed	31-July-2015
Vial Label	SC11-016 p33+9 31July2015 RAS
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

Testing Performed by WiCell

Test Description	Test Provider	Test Method	Test Specification	Result
Karyotype by G-banding	WiCell	SOP-CH-003	Expected karyotype	See Report
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	Recoverable attachment after passage	Pass
Identity by STR	UW Translational Research Initiatives in Pathology Laboratory	PowerPlex 16 HS System by Promega	Consistent with STR profile of deposited cell line	Pass
Sterility	Steris	ST/07	Negative	Pass
Mycoplasma	WiCell	SOP-QU-004	Negative	Pass

Testing Reported by Provider

The Provider stated that some or all of the additional analyses listed below may have been performed for this cell line. For more information, publication and Synapse links, where available, are provided on the cell line specific web page on the WiCell website.

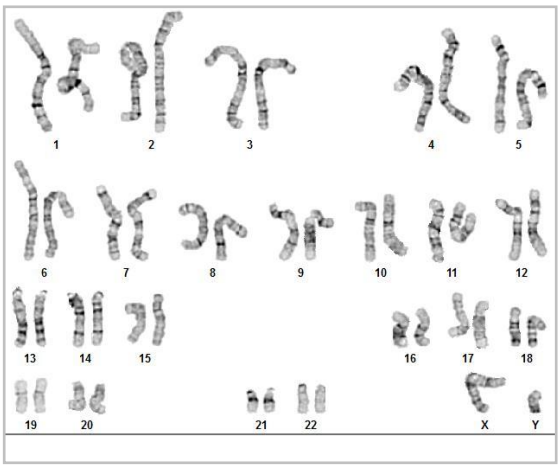
- RNA-Seq
- Teratoma representative of all three embryonic germ layers identified in all tumors with histopathological analysis
- SNP microarray
- Immunostaining analysis to confirm pluripotency and OCT4 to evaluate the presence of undifferentiated PSC
- mRNA, miRNA, and methylation profiling
- Genemoics characterization
- Flow Cytometry (SSEA-1, SSEA-4, Tra 1-61,Tra 1-80, CD9, OCT-4)

Approval Date	Quality Assurance Approval
17-January-2019	9/4/2020 X HEB HEB Quality Assurance Signed by: Bruner, Haley

Chromosome Analysis Report: 074323

Date Reported: Monday, December 31, 2018
Cell Line: iPS DF19-9-7T-DB36826 14200
Passage#: 34
Date of Sample: 12/18/2018
Specimen: Human IPS
Results: 46,XY

Cell Line Sex: Male
Reason for Testing: Lot Release Testing
Investigator: [redacted], WiCell



Cell: 3
Slide: G03
Slide Type: Karyotype

Total Counted: 20
Total Analyzed: 8
Total Karyogrammed: 4
Band Resolution: 425 - 550

Interpretation:

This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.

Completed by: [redacted], CG(ASCP)
Reviewed and Interpreted by: [redacted], PhD, FACMG

Date:_____ **Sent By:**____ **Sent To:**_____ **QC Review By:** ____

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e.,mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

Short Tandem Repeat Analysis

TRIPath
HISTOLOGY - IHC - MOLECULAR - IMAGING
Department of Pathology and Laboratory Medicine
TRIP Laboratory (Molecular)
https://research.pathology.wisc.edu/trip/
(608) 265-9168

WiCell
Your Lab Partner
characterization@wicell.org
(608) 316-4145

Sample Report:
14200-STR
Sample Name on Tube: 14200-STR
79.1 ng/µL, (A260/280=1.89)
Sample Type: Cells
Cell Count: ~2 million cells

Requestor:
WiCell Research Institute
Quality Assurance Department

Receive Date: 01/02/19
Report Sent: 01/07/19
Assay Date: 01/02/19
File Name: STR 190103 revised wmr
Report Date: 01/07/19

STR Locus	STR Genotype Repeat #	STR Genotype
FGA	16–18,18.2,19,19.2,20,20.2,21,21.2,22, 22.2, 23, 23.2, 24, 24.2, 25, 25.2, 26–30, 31.2, 43.2, 44.2,45.2, 46.2	Identifying information has been redacted to protect donor confidentiality. If more information is required, please, contact WiCell's Technical Support.
TPOX	6-13	
D8S1179	7-18	
vWA	10-22	
Amelogenin	X,Y	
Penta_D	2.2, 3.2, 5, 7-17	
CSF1PO	6-15	
D16S539	5, 8-15	
D7S820	6-14	
D13S317	7-15	
D5S818	7-16	
Penta_E	5-24	
D18S51	8-10, 10.2, 11-13, 13.2, 14-27	
D21S11	24,24.2,25,25.2,26-28,28.2,29,29.2, 30, 30.2,31, 31.2,32,32.2,33,33.2, 34,34.2,35,35.2,36-38	
TH01	4-9,9.3,10-11,13.3	
D3S1358	12-20	

Results: **Based on the 14200-STR cells submitted by WiCell QA dated and received on 01/02/19, this sample (Label on Tube: 14200-STR) exactly matches the STR profile of the human stem cell line iPS DF19-9-7T comprising 27 allelic polymorphisms across the 15 STR loci analyzed.**

Interpretation: **No STR polymorphisms other than those corresponding to the human iPS DF19-9-7T stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggests that the 14200-STR sample submitted corresponds to the iPS DF19-9-7T stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells.**

Sensitivity: **Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~2-5%.**

X *RMB* **Digitally Signed on 01/07/19**

[redacted], *BA*
TRIP Laboratory, Molecular

X *WMR* **Digitally Signed on 01/07/19**

[redacted], *PhD, Director / Co-Director*
UWHC Molecular Diagnostics Laboratory / UWSMPH TRIP Laboratory

Native Product Sterility Report

STERIS

SAMPLE #:	18121076
DATE RECEIVED:	13-Dec-18
TEST INITIATED:	17-Dec-18
TEST COMPLETED:	31-Dec-18

WiCell
504 S Rosa Road, Rm 101
Madison, WI 53719

SAMPLE NAME / DESCRIPTION: LUEL8363i-3 WB66952 14169, JHU088i DB41240 14170, JHU132i DB36272 14171, JHU102i DB41279 14172, JHU103i DB36236 14173, iPS DF19-9-7T DB36826 14174, iPS DF4-3-7T.A DB36837 14175, iPS DF6-9-9T.B DB36844 14176, MIRJT7i-mND2-0 DB36854 14177, IISH1i-BM1 DB36864 14178, IISH2i-BM9 DB36871 14179, IISH3i-CB6 DB36881 14180, WA01 DB36785 14181, WA07 DB36888 14182, WA09 DB36899 14183, WA24 DB36798 14184, JHU155i DB41365 14185, JHU184i DB41392 14186, JHU142i DB41344 14187, WA09 NA65899 14189.

UNIQUE IDENTIFIER: NA

TEST RESULTS:

# Tested	# Positives (Growth)	- Control
20	1	2 Negatives

TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
20	TSB	40	20-25	14
20	FTG	40	30-35	14

REFERENCE: Processed according to LAB-003: Sterility Test Procedure

PD #: 000053

TEST METHODOLOGY: USP - Direct Transfer

COMMENTS: Sample labeled P142 P6 6/16/16 0.9M is positive in TSB and FTG media.

Reported as per packing slip.

REVIEWED BY _____ DATE 02JAN19

Mycoplasma Detection Assay Report

Testing Performed by WiCell

Lot Release Testing

December 13, 2018

FORM SOP-QU-004.01
Version G Edition 02
Reported by: SM
Reviewed by: JB
Berthold Flash n' Glow 539

#	Sample Name	Reading A		A Ave	Reading B		B Ave	Ratio B/A	Result	Comments/Suggestions
		RLU1	RLU2		RLU1	RLU2				
1	iPS DF19-9-7T-DB36826 14200	108	102	105	49	49	49	0.47	Negative	
2	Positive (+) Control	190	185	187.5	10777	10762	10770	57.44	Positive	
3	Negative (-) Control	297	292	294.5	29	26	27.5	0.09	Negative