

Cell Characterization

Guidelines for characterization testing of cell lines

Assay	What it Detects	What it Doesn't Detect	TATi	When to Use
Fluorescence In Situ Hybridization (FISH)	Genomic sequence of interest Duplications or deletions >100 Kb >2% mosaicism (for example: cultures where >2 of 100 cells are trisomy 12) Chromosomal location of genomic gains Chromosome fusions (breakaparts)	Changes in regions other than the probespecific sequence	10-15 days	To confirm findings and refine breakpoints detected by g-banded karyotyping To confirm findings and localize genomic gains detected by SNP microarray As a screen for microdeletions/duplications of known targets
G-Banded Karyotyping	Microscopic genomic abnormalities (>5-10 Mb) Inversions Duplications/deletions Balanced and unbalanced translocations Aneuploidies >10% mosaicism (for example: cultures where >1 of 10 cells are trisomy 12)	Submicroscopic genomic abnormalities (<5 Mb) <p><~10% culture mosaicism (for example: cultures where 1 of 10 cells is trisomy 12)</p>	7-10 days (4-6 days expedited)	As a baseline genomic screen At derivation of cell lines At the start of experimental protocols To assess and monitor genomic stability (for example: every 5-10 passages of cell culture) At conclusion of experiments (prior to publication) For cell line banking When publication-quality karyotypes are needed
Mycoplasma Detection by PCR	96 species of mycoplasma contamination from stem cell cultures. Sensitivity (5-100 CFU/ml)	This system does not allow for the amplification of DNA originating from other sources, such as bacteria.	5-7 days	To monitor the health of your cell line To monitor for contamination in shared lab spaces To assure that mycoplasma is not interfering with your experiments To rule out mycoplasma as the culprit of chromosomal aberrations
Short Tandem Repeat Analysis (STR)	STR polymorphisms for 15 loci plus amelogenin (Promega® PowerPlex® 16) Probability of matching identity to an existing STR profile	STR polymorphisms in areas other than those represented in Promega® PowerPlex® 16	10-20 days	To monitor identity of a cell line To confirm relationship of iPS cells to their parent line To establish an STR profile of a newly derived or reprogrammed cell line To rule out culture cross-contamination
Single nucleotide polymorphism (SNP) Microarray	Submicroscopic genomic abnormalities (>5- 10 Mb) Genomic gains and losses (>50 Kb) Copy number variants Duplications/deletions Unbalanced translocations Aneuploidies Copy neutral Loss of Heterozygosity (LOH) / Absence of Heterozygosity (AOH) (>5 Mb) >~10% mosaicism (for example: cultures where >1 of 10 cells are trisomy 12	Balanced translocations Robertsonian Balanced insertions Inversions < ~10% culture mosaicism (for example: cultures where 1 of 10 cells is trisomy 12) Chromosomal position of genomic gains	14-21 days	As a baseline genomic screen To detect submicroscopic (<5 Mb) abnormalities To identify amplified or deleted genes of interest To assess and monitor genomic stability (for example: every 5-10 passages of cell culture) In conjunction with G-banded karyotyping To define unbalanced translocation breakpoints For research of genomic copy number change To identify structural variation within populations or disease cohorts To develop a cell line copy number variant profile
Spectral Karyotyping (SKY)	Microscopic genomic abnormalities (>5-10 Mb) Balanced and unbalanced translocations Aneuploidies	Submicroscopic genomic abnormalities (<5 Mb) Inversions Duplications/deletions	14-21 days	As an adjunct to g-banded karyotyping To define complex rearrangements To identify marker chromosomes When publication-quality spectral karyotypes are needed

¹ Turn-around-times (TAT) provided are based on provision of sufficient mitotically active hES or iPS cultures grown in Matrigel/TeSR or MEF/hES media conditions.



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