

Thaw and Culture Details

Cell Line Name	STAN009i-165-2						
WiCell Lot Number	DB31110						
Provider	Stanford University – Laboratory of Dr. Marlene Rabinovitch						
Banked By	Stanford University – Laboratory of Dr. Marlene Rabinovitch						
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 3 wells of a 6 well plate. WiCell recommends thawing using ROCK Inhibitor for best results.						
	Feeder Independent						
	Medium: E8						
	Matrix: Matrigel®						
Protocol	WiCell Feeder Independent E8 Medium Protocol						
Passage Number	p10 These cells were cultured for 10 passages prior to freeze and post reprogramming. Add +1 to the passage number to best represent the overall passage number of the cells at thaw.						
Date Vialed	10-June-2015						
Vial Label	06/10/2015 E 165 D####-### ip 165FSVNOC2 P10 V####################################						
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.						

Testing Performed by WiCell

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Test Description	Test Provider	Test Method	Test Specification	Result					
Karyotype by G-banding	WiCell	SOP-CH-003	Expected karyotype	Pass					
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	Recoverable attachment after passage	Pass					
Identity by STR	UW Translational Research Initiatives in Pathology Laboratory	PowerPlex 16 HS System by Promega	Defines profile	Pass					
Sterility	Steris	ST/07	Negative	Pass					
Mycoplasma	WiCell	SOP-QU-004	Negative	Pass					

Testing Reported by Provider

Test Description	Method	Result
Identity	SNP	iPSCs match the donor material
Mycoplasma	Lonza MycoAlert™ kit	Negative

The Provider stated that the additional analysis listed below may have been performed for this cell line. For more information, publication and dbGaP links, where available, are provided on the cell line specific web page on the WiCell website.

Infinium® Expanded Multi-Ethnic Genotyping Array (MEGAEX)



Approval Date	Quality Assurance Approval			
	9/21/2017			
04-June-2016	X RK			
	RK			
	Quality Assurance			
	Signed by: Kremers, Erik			



Chromosome Analysis Report: 067691

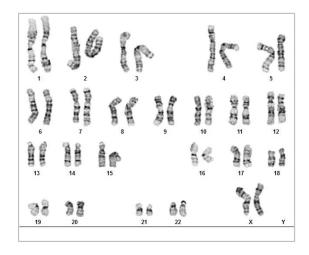
Date Reported: Monday, September 11, 2017

Cell Line: STAN009i-165-2-DB31110 12756

Passage#: 12

Date of Sample: 8/27/2017 Specimen: Human IPSC

Results: 46,XX



Cell Line Gender: Female

Reason for Testing: lot release testing

Investigator:

Cell: 42

Slide: G01

Slide Type: Karyotype

Total Counted: 20 Total Analyzed: 8

Total Karyogrammed: 4 Band Resolution: 400 - 450

QC Review By: __

Interpretation:

Date:

This is a normal karyotype. No clonal abnormalities were detected at the stated band level of resolution.

Completed by:

Reviewed and Interpreted by:

cell populations in this specimen (i.e.,mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

A signed copy of this report is available upon request.

Sent By:____ Sent To:_ Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal

This assay was conducted solely for listed investigator/institution. The results may not be relied upon by any other party without the prior written consent of the Director of the WiCell Cytogenetics Laboratory. The results of this assay are for research use only. If the results of this assay are to be used for any other purpose, contact the Director of the WiCell Cytogenetics Laboratory.

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Short Tandem Repeat Analysis

WiCell®
info@wicell.org
(888) 204-1782

Department of Pathology and Laboratory Medicine TRIP Laboratory (Molecular)

http://www.pathology.wisc.edu/research/trip

Sample Report: 12756-STR

Sample Name on Tube: 12756-STR

 $72.9 \text{ ng/}\mu\text{L}$, (A260/280=2.13)

Sample Type: Cells

Cell Count: ~2 million cells

Requestor:

WiCell Research Institute
Ouality Department

Sample Date: N/A **Receive Date:** 09/05/17

Assay Date: 09/12/17 File Name: 170913 STR WMR

Report Date: 09/15/17

STR Locus	STR Genotype Repeat #	STR Genotype				
FGA	16–18,18.2,19,19.2,20,20.2,21,21.2,22, 22.2, 23, 23.2, 24, 24.2, 25, 25.2, 26–30, 31.2, 43.2, 44.2,45.2, 46.2	Identifying information has				
TPOX	6-13	been redacted to protect donor				
D8S1179	7-18					
vWA	10-22	confidentiality. If				
Amelogenin	X,Y	more information				
Penta_D	2.2, 3.2, 5, 7-17	is required, please, contact				
CSF1PO						
D16S539	5, 8-15	WiCell's Technica Support.				
D7S820	6-14					
D13S317	7-15					
D5S818	7-16					
Penta E	5-24					
D18S51	8-10, 10.2, 11-13, 13.2, 14-27					
D21S11	24,24.2,25,25.2,26-28,28.2,29,29.2, 30, 30.2,31, 31.2,32,32.2,33,33.2, 34,34.2,35,35.2,36-38					
TH01	4-9,9.3,10-11,13.3					
D3S1358	12-20					

<u>Results:</u> Based on the 12756-STR cells submitted by WiCell QA dated and received on 09/05/17, this sample (Label on Tube: 12756-STR) defines the STR profile of the human stem cell line STAN009i-165-2 comprising 28 allelic polymorphisms across the 15 STR loci analyzed.

<u>Interpretation:</u> No STR polymorphisms other than those corresponding to the human STAN009i-165-2 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggests that the 12756-STR sample submitted corresponds to the STAN009i-165-2 stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells.

<u>Sensitivity:</u> Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~2-5%.

X RMB	Digitally Signed on 09/15/17	X WMR	Digitally Signed on	09/15/17
TRIP La	boratory, Molecular	UWHC Mol	, PhD, Director / Co-Director ecular Diagnostics Laboratory / UWSI	

Native Product Sterility Report



SAMPLE #:

17081530

DATE RECEIVED:

24-Aug-17

TEST INITIATED:

25-Aug-17

TEST COMPLETED:

08-Sep-17

SAMPLE NAME / DESCRIPTION:

WA09-RB66490 12757

WA09-RB66492 12758

WA09-RB66493 12759

WC026i-5807-3 WB66538 12760 WC032i-6007-1-WB66540 12761 WC033i-6007-2-WB66539 12762 STAN008i-165-1-DB31100 12763 STAN009i-165-2-DB31110 12764 STAN053i-149-1-DB30924 12765 STAN001i-047-1-DB31118 12766

UNIQUE IDENTIFIER:

NΔ

PRODUCT REGISTRATION:

Human iPS cells

TEST RESULTS:

WiCell

504 S Rosa Rd, Rm 101

Madison, WI 53719

	# Positives	
# Tested	(Growth)	- Control
10	0	2 Negatives

TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
10	TSB	40	20 - 25	14
10	FTG	40	30 - 35	14

REFERENCE:

Processed according to LAB-003: Sterility Test Procedure

METHOD VALIDATION / PD #:

000053

TEST METHODOLOGY:

USP - Direct Transfer

Native Product Sterility Report



COMMENTS:

Sample # 17081530

REVIEWED BY Dessone

DATE USEPIT

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests.



Mycoplasma Detection Assay Report Testing Performed by WiCell

Testing Performed by WiCell Lot Release Testing August 28, 2017

FORM SOP-QU-004.01 Version F Edition 02 Reported by: KR Reviewed by: JB Berthold Flash n' Glo 539

		Reading A		A	Read	ling B	В	Ratio		
#	Sample Name	RLU1	RLU2	Ave	RLU1	RLU2	Ave	B/A	Result	Comments/Suggestions
1	STAN009i-165-2-DB31110 12756	107	101	104	27	24	25.5	0.25	Negative	
2	Positive (+) Control	165	166	165.5	17611	17643	17627	106.51	Positive	
3	Negative (-) Control	257	263	260	36	30	33	0.13	Negative	

