

Certificate of Analysis - Amended

Product Description	WA09 Distribution Lot	WA09 Distribution Lot			
Cell Line Provider	WiCell research Institute	WiCell research Institute			
Parent Material	WA09-MCB-01	WA09-MCB-01			
Lot Number	WA09-DL-08	WA09-DL-08			
Date Vialed	15-December-2008	15-December-2008			
Passage Number	P25	P25			
Culture Platform	Feeder Dependent - MEFs	Feeder Dependent - MEFs			
	Media: hES Medium	Media: hES Medium Matrix: MEFs			

The following testing specifications have been met for the specified product lot:

Test Description	Test Provider	Test Method	Test Specification	Result
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	≥ 15 Undifferentiated Colonies, ≤ 30% Differentiation	Pass
Identity by STR	UW Molecular Diagnostics Laboratory	PowerPlex 1.2 System by Promega	Positive Identity	Pass
Sterility - Direct transfer method	Apptec	30744	No contamination detected	Pass
Mycoplasma	Bionique	M250	No contamination detected	Pass
Karyotype by G-banding	WiCell	SOP-CH-003	Normal karyotype	Pass
Flow Cytometry for ESC Marker Expression	UW Flow Cytometry Laboratory	SOP-CH-101 SOP-CH-102 SOP-CH-103 SOP-CH-105	Report - no specification	See report

Distribution Lot cells are expanded from vials of Master Cell Bank (MCB) cells. MCB cells are thoroughly tested and known to be free of many viruses and pathogens. These cells have undergone extensive testing and are not known to harbor any human pathogens or adventious agents of murine, bovine, or porcine origin. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells.

Please contact technical service via the website to request test methods and other assistance with your cells. The knowledgeable technical support staff can assist with cell culture concerns, training, and any other customer service concerns.

Amendment(s):

Reason for Amendment	Date
CoA updated to include copyright information and update logo.	See signature
CoA updated for format changes, clarification of test specifications, test method, addition of test provider, culture platform, and electronic signature, and reference to WiCell instead of the NSCB	17-AUG-2010
Original CoA	17-MAR-2009

Date of Lot Release	Quality Assurance Approval
17-March-2009	1/3/2014 X AMC
11 march 2000	AMC Quality Assurance Signed by:

©2009 WiCell Research Institute The material provided under this certificate has been subjected to the tests specified and the results and data described herein are accurate based on WiCell's reasonable knowledge and belief. Appropriate Biosafety Level practices and universal precautions should always be used with this material. For clarity, the foregoing is governed solely by WiCell's Terms and Conditions of Service, which can be found at http://www.wicell.org/privacyandterms.





Short Tandem Repeat Analysis*

Sample Report: 2988-STR

UW HLA#: 60469

Sample Date: 03/04/09

Received Date: 03/04/09

Requestor: WiCell Research Institute

Test Date: 03/12/09

File Name: 090313

Report Date: 03/16/09

Sample Name: (label on tube) 2988-STR

Description: DNA Extracted by WiCell

251.30 ug/mL; 260/280 = 1.89

Locus	Repeat #	STR Genotype
D16S539	5, 8-15	12,13
D7S820	6-14	9,11
D13S317	7-15	9,9
D5S818	7-15	11,12
CSF1PO	6-15	11,11
TPOX	6-13	10,11
Amelogenin	NA	X,X
TH01	5-11	9.3,9.3
vWA	11, 13-21	17,17

Comments: Based on the DNA 2988-STR dated 03/04/09 and received on 03/04/09 from WI Cell, this sample (UW HLA# 60469) matches exactly the STR profile of the human stem cell line H9 comprising 12 allelic polymorphisms across the 8 STR loci analyzed. No STR polymorphisms other than those corresponding to the human H9 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggest that the 2988-STR DNA sample submitted corresponds to the H9 stem cell line and it was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells. Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~5%.

	Data	Data
68	Date	Date
HLA/Molecular Diagnostics Laboratory		HLA/Molecular Diagnostics Laboratory

File: Final STR Report

^{*} Testing to assess engraftment following bone marrow transplantation was accomplished by analysis of human genetic polymorphisms at STR loci. This methodology has not yet been approved by the FDA and is for investigational use only.

This report is confidential. No part may be used for advertising or public announcement without written permission. Results apply only to the sample(s) tested.



WiCell Research Institute

Report Number 800061 Page 1 of 1

February 13, 2009 P.O. #:

STERILITY TEST REPORT

Sample Information:

hES Cells, WA09-DL-8

Date Received:
Date in Test:

January 27, 2009

Date in Test:

Date Completed:

January 30, 2009 February 13, 2009

Test Information:

Test Codes: 30744, 30744A Immersion, USP / 21 CFR 610.12 Procedure #: BS210WCR.201

TEST PARAMETERS	PRODUCT		
Approximate Volume Tested	0.5 mL	0.5 mL 2 FTM 400 mL 14 Days 30 °C to 35 °C	
Number Tested	2		
Type of Media	SCD		
Media Volume	400 mL		
Incubation Period	14 Days		
Incubation Temperature	20 °C to 25 °C		
RESULTS	2 NEGATIVE	2 NEGATIVE	

QA Reviewed:

02-16-09

Reviewed:

02-13-09



BIONIOUE TESTING LABORATORIES, INC

APPENDIX I		
Document #: Edition #: Effective date:	DCF3008A 06 9/17/2003	
Title:	DNA FLUOROCHROME ASSAY RESULTS	
	DNA-FLUOROCHROME ASSAY RESULTS Procedures 3008, 3009, 3011	
Sample ID # <u>56324</u>	M-250 Date Rec'd: 02/10/2009 P.O. #	
Indicator Cells Inoculated:	Date/Initials: 2/12/09 / JA	
Fixation:	Date/Initials: Z 16 09 / JA	
Staining;	Date/Initials: $2/16/09/5\%$	
TEST/CONTROL ARTICLE:		
WA09-DL-8		
LOT# <u>NA</u>		
Wicell QA WiCell Research Instit	<u>ute</u>	
,		
DNA FLUOROCHROME A	SSAY RESILTS:	
NEGATIVE:	A reaction with staining limited to the nuclear region, which indicates	
THE CETTE VE	no mycoplasmal contamination.	
POSITIVE:	A significant amount of extranuclear staining which strongly suggests mycoplasmal contamination.	;
INCONCLUS	IVE:	
	A significant amount of extranuclear staining consistent with low-level mycoplasmal contamination or nuclear degeneration.	
	A significant amount of extranuclear staining consistent with bacterial, fungal or other microbial contaminant or viral CPE. Microbiology not consistent for mycoplasmal contamination.	
COMMENTS:		
Date: Z/16/09 Results	Read by: JA Date of Review: 2-17-09 Reviewed by: 59M	į.





APPENDIX IV

Page 1 of 2

Document#: Edition#:

DCF3013D

10

Effective Date:

07/15/2003

Title:

M-250 FINAL REPORT SHEET

M-250 FINAL REPORT

Direct Specimen Culture Procedure 3008, 3011, 3013

TO: Wicell QA

WiCell Research Institute

BTL SAMPLE ID#: 56324

P.O.#:

DATE REC'D:

02/10/2009

TEST/CONTROL ARTICLE:

WA09-DL-8

LOT#: NA

DIRECT CULTURE SET-UP (DAY 0)	DA	ATE:	02/11/200	9
INDICATOR CELL LINE (VERO)	SEE DNA FLUO	ROCHRO	DME RECORD SHEET	
	8			DATE
THIOGLYCOLLATE BROTH	DAY 7	+	Θ	02/18/2009
	DAY 28	+	\odot	03/11/2009
BROTH-FORTIFIED COMMERCIAL				
0.5 ml SAMPLE	DAY 7	+	0	02/18/2009
6.0 mL BROTH	DAY 28	+	\odot	03/11/2009
BROTH-MODIFIED HAYFLICK				
0.5 mL SAMPLE	DAY 7	+	Θ	02/18/2009
6.0 mL BROTH	DAY 28	+	\odot	03/11/2009
BROTH-HEART INFUSION				
0.5 mL SAMPLE	DAY 7	+	Θ	02/18/2009
6.0 mL BROTH	DAY 28	+	(03/11/2009
(See Reverse)		6		

Document#:

DCF3013D

Edition#:

10

Effective Date:

07/15/2003

Title:

M-250 FINAL REPORT SHEET

SAMPLE ID#: 56324		AEROBIC	MICROAEROPHILIC	DATE
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7 DAY 14 DAY 21	+ () + () + ()	+ (-) + (-)	$\frac{02/18/2009}{02/25/2009}$ $\frac{03/04/2009}{03}$
AGAR PLATES-MODIFIED HAYFLICK	DAY 7 DAY 14 DAY 21	+ (-) + (-) + (-)	+ (-) + (-)	$\frac{02/18/2009}{02/25/2009}$ $\frac{03/04/2009}{03/04/2009}$
AGAR PLATES-HEART INFUSION	DAY 7 DAY 14 DAY 21	+ (D) + (D) + (D)	+ (-) + (-) + (-)	$\frac{02/18/2009}{02/25/2009}$ $\frac{03/04/2009}{03}$
BROTH SUBCULTURES (DAY 7)		DATE: <u>02/</u>	18/2009	
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7 DAY 14 DAY 21	+ (5) + (6) + (7)	+ (=) + (=) + (=)	$\frac{02/25/2009}{03/04/2009}$ $\frac{03/11/2009}{03/11/2009}$
AGAR PLATES-MODIFIED HAYFLICK	DAY 7 DAY 14 DAY 21	+ (D) + (D) + (D)	+ (-) + (-) + (-)	$\frac{02/25/2009}{03/04/2009}$ $\frac{03/11/2009}{03/11/2009}$

RESULTS:

No detectable mycoplasmal contamination

3-11-09

Date

M-250 Procedural Summary: The objective of this test is to ascertain whether or not detectable mycoplasmas are present in an in vitro cell culture sample, be it a primary culture, hybridoma, master seed stock or cell line. This procedure combines an indirect DNA staining approach to detect non-cultivable mycoplasmas with a direct culture methodology utilizing three different mycoplasmal media formulations. The indirect approach involves the inoculation of the sample into a mycoplasma-free VERO (ATCC) indicator cell line and performing a DNA fluorochrome assay after 72-120 hours of incubation. The direct culture aspect of the test utilizes three different mycoplasmal media including both broth and agar formulations. The sample is inoculated into each of the 3 broth formulations and also onto duplicate plates (0.1 mL/plate) for each of the 3 agar formulations. Subculture from broth to fresh agar plates is carried out after 7 days incubation. Agar plates are incubated aerobically and microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final microaerophillically indicative of mycoplasmal contamination.



WiCell Cytogenetics Report: 000915-020909

NSCB 2988

Report Date: February 18, 2009

Case Details:

Cell Line: WA09-DL-8 (2988)

Passage #: 30

Date Completed: 2/18/2009
Cell Line Gender: Female

Investigator: National Stem Cell Bank

Specimen: hESC on MEF feeder

Date of Sample: 2/9/2009

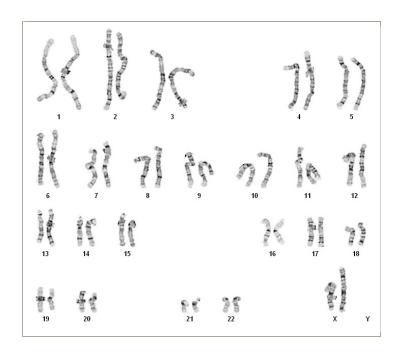
Tests, Reason for: NSCB- DL Testing

Results: 46,XX

Completed by CLSp(CG), on 2/13/2009

Reviewed and interpreted by PhD, FACMG, on 2/18/2009

Interpretation: No clonal abnormalities were detected at the stated band level of resolution.



Cell: S01-03

Slide: B

Slide Type: Karyotyping

Cell Results: Karyotype: 46,XX

of Cells Counted: 20

of Cells Karyotyped: 4

of Cells Analyzed: 8

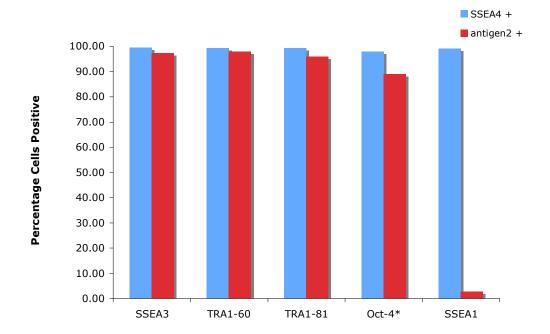
Band Level: 475-675

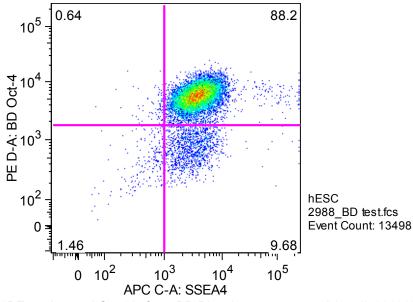
Results Transmitted by Fax / Email / Post Sent By:

Date:_____Sent To:

Procedures performed: SOP-CH-101 SOP-CH-102 SOP-CH-103 SOP-CH-105 Cell Line: Passage Sample ID: 2988-FAC **Date of**: (mm/dd/yy) acquisition: 2/10/09 file creation: 2/27/09 file submission: 2/27/09

	SSEA4 -	SSEA4 +	SSEA4+	SSEA4 -	ALL	ALL
	22211.				1122	1122
<u>antigen2:</u>	<u>antigen2 +</u>	<u>antigen2 +</u>	<u>antigen2 -</u>	<u>antigen2 -</u>	<u>SSEA4 +</u>	<u>antigen2 +</u>
SSEA3	0.01	97.20	2.16	0.62	99.36	97.21
TRA1-60	0.44	97.40	1.76	0.37	99.16	97.84
TRA1-81	0.27	95.70	3.57	0.45	99.27	95.97
Oct-4*	0.64	88.20	9.68	1.46	97.88	88.84
SSEA1	0.14	2.49	96.60	0.77	99.09	2.63





*PE-conjugated Oct-3/4 from BD Biosciences was used (cat #560186).