Certificate of Analysis

Product Description	WA09	
Cell Line Provider	WiCell Research Institute	
Parent Material	WA09-MCB-01	
Lot Number	WB0007	
Date Vialed	22-April-2010	
Passage Number	p22[1]	
Culture Platform	Feeder Independent	
	Media: TeSR	Matrix: Matrigel

The following testing specifications have been met for the specified product lot:

Test Description	Test Provider	Test Method	Test Specification	Result
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	≥ 15 Undifferentiated Colonies, ≤ 30% Differentiation	Pass
Identity by STR	UW Molecular Diagnostics Laboratory	PowerPlex 1.2 System by Promega	Consistent witih known profile	Pass
Sterility - Direct transfer method	Apptec	30744	Negative	Pass
Mycoplasma	Bionique	M250	No contamination detected	Pass
Karyotype by G-banding	WiCell	SOP-CH-003	Normal karyotype	Pass

[1]Due to space limitations, only the overall passage number is included on the vial. Prior to freeze, these cells were cultured for a total of 21 passages, 5 of them in mTeSR1/Matrigel. WiCell adds +1 to the overall passage number at freeze so that the number on the vial best represents the overall passage number of the cells at thaw. Foornote provided by T.L. 29Oct10.

Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells.

Amendment(s):

Reason for Amendment	Date
CoA updated to include copyright information.	See signature
CoA updated for clarification of passge number, test specifications, product description, and removed text regarding technical services and distribution lots	01-November-2010
Original CoA	12-August-2010

Date of Lot Release	Quality Assurance Approval
12-August-2010	1/3/2014 X AMC AMC Quality Assurance Signed by [redacted]

UW Health

University of Wisconsin
Hospital and Clinics

Histocompatibility/Molecular Diagnostics Laboratory

Short Tandem Repeat Analysis*

Sample Report: 5771-STR | UW HLA#: 63399 | Sample Date: 07/02/10
Received Date: 07/02/10

Requestor: WiCell Research Institute

Test Date: 07/06/10 | File Name: 100707 | Report Date: 07/12/10

Sample Name: (label on tube) **5771-STR**

Description: DNA Extracted by WiCell
250.37 ug/mL; 260/280 = 1.90

Locus	Repeat #	STR Genotype
D16S539	5, 8-15	12,13
D7S820	6-14	9,11
D13S317	7-15	9,9
D5S818	7-15	11,12
CSF1PO	6-15	11,11
TPOX	6-13	10,11
Amelogenin	NA	X,X
TH01	5-11	9.3,9.3
vWA	11, 13-21	17,17

Comments: Based on the DNA 5771-STR dated and received on 07/02/10 from WI Cell, this sample (UW HLA# 63399) matches exactly the STR profile of the human stem cell line H9 comprising 12 allelic polymorphisms across the 8 STR loci analyzed. No STR polymorphisms other than those corresponding to the human H9 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggest that the 5771-STR DNA sample submitted corresponds to the H9 stem cell line and it was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells. Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is estimated to be ~5%.

1/13/10
Date
HLA/Molecular Diagnostics Laboratory

07/13/10
Date
HLA/Molecular Diagnostics Laboratory

* Testing to assess engraftment following bone marrow transplantation was accomplished by analysis of human genetic polymorphisms at STR loci. This methodology has not yet been approved by the FDA and is for investigational use only.

File: Final STR Report

Test Facility:

WuXi AppTec

Report Number
837137

WiCell Research Institute

June 02, 2010
P.O. #:

STERILITY TEST REPORT

Sample Information:
hES Cells
1: WA09-WB0007 # 5170
2: WA18-WB0003 # 0651
3: WA18-WB0010 # 8027
4: WA19-WB0015 # 7336
5: WA19-WB0013 # 5777
6: WA20-WB0014 # 9912
7: iPS(IMR90)-3-MCB-01 #3377
8: iPS(Foreskin)-3-WB0002 # 2503

Date Received: May 13, 2010
Date in Test: May 18, 2010
Date Completed: June 01, 2010

Test Information:
Test Codes: 30744, 30744A
Immersion, USP / 21 CFR 610.12
Procedure #: BS210WCR.201

TEST PARAMETERS	PRODUCT	
Approximate Volume Tested	0.5 mL	0.5 mL
Number Tested	16	16
Type of Media	SCD	FTM
Media Volume	400 mL	400 mL
Incubation Period	14 Days	14 Days
Incubation Temperature	20 °C to 25 °C	30 °C to 35 °C
RESULTS	16 NEGATIVE	16 NEGATIVE

QA Reviewer 06-03-10 **Date**

Technical Reviewer 06-02-10 **Date**

Testing conducted in accordance with current Good Manufacturing Practices.

APPENDIX

Document ID #:	DCF9002F
Title:	QUALITY ASSURANCE REPORT - GMP
Effective Date:	03/12/10
Edition #:	01

QUALITY ASSURANCE REPORT – GMP

TEST PERFORMED	PROCEDURAL REFERENCE	TEST PERFORMED	PROCEDURAL REFERENCE
☒ M-250	SOP's 3008, 3011, 3013	☐ M-700	SOP's 3008, 3009, 3010
☐ M-300	SOP's 3008, 3014	☐ M-800	SOP's 3008, 3011, 3016
☐ M-350	SOP's 3008, 3014, 3015		

Bionique Sample ID #(s) 61671

This testing procedure was performed in compliance with the FDA's Current Good Manufacturing Practice (cGMP) standards (to the extent that the regulations pertain to the procedures performed) as specified in the Code of Federal Regulations, Title 21 Parts 210 and 211 [21 CFR 210 & 211]. All related records derived from the test procedures have been reviewed by the Quality Assurance Department. The individual's signature below verifies that the methods and procedures referenced above have been followed and that the Final Report accurately reflects the raw data generated during the course of the procedures. All records, including raw data and final reports are archived on site for a minimum of seven years.

The specified test's procedures determine the intervals at which samples are inspected. The medium used for testing must pass quality control mycoplasmal growth promotion testing and sterility testing. Traceability of all of the components used is assured and supporting documentation can be supplied upon request.

Quality Assurance Review Date: 7/28/10

Reviewed By , QA Assistant: ___

NOTE:

1. Prior to receipt at Bionique® Testing Laboratories, Inc., the stability of the test article is the responsibility of the company submitting the sample. Bionique Testing Laboratories Inc. will assume responsibility for sample stability following receipt and prior to being placed on test.
2. This test is for the detection of microbiological growth and does not require statistical validation.

REFERENCES

Regulatory:

1. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Code of Federal Regulations [CFR], Title 21 CFR Part 210, Current Good Manufacturing Practice in Manufacturing, Processing, Packing, or Holding of Drugs; General. FDA. Office of the Federal Register, National Archives and Records Department.
2. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Code of Federal Regulations [CFR], Title 21 CFR Part 211, Current Good Manufacturing Practice for Finished Pharmaceuticals. FDA. Office of the Federal Register, National Archives and Records Department.
3. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Points to Consider in the Characterization of Cell Lines Used to Produce Biologicals, Director, Center for Biologics Evaluation and Research, FDA. May, 1993. Docket No. 84N-0154.
4. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Code of Federal Regulations [CFR], Title 21 CFR Part 610.30, General Biological Products Standards; Subpart D, Test for Mycoplasma. FDA. Office of the Federal Register, National Archives and Records Department.

General:

1. Barile MF, Kern J. Isolation of Mycoplasma arginini from commercial bovine sera and its implication in contaminated cell cultures. Proceedings of the Society for Experimental Biology and Medicine, Volume 138, Number 2, November 1971.
2. Chen, T.R. In situ detection of mycoplasma contamination in cell cultures by fluorescent Hoechst 33258 stain. Experimental Cell Research, 104: 255-262, 1977.
3. Carolyn K. Lincoln and Daniel J. Lundin. Mycoplasma Detection and Control. U. S. Fed. for Culture Collections Newsletter, Vol. 20, Number 4, 1990.
4. Fetal Bovine Serum; Proposed Guideline. National Committee For Clinical Laboratory Standards (NCCLS), Vol. 10, Number 6, 1990. (NCCLS publication M25-P).
5. McGarrity GJ, Sarama J, Vanaman V. Cell Culture Techniques. ASM News, Vol. 51, No. 4, 1985.
6. Tully JG, Razin S. Methods in Mycoplasmology, Volumes I and II. Academic Press, N.Y., 1983.
7. Barile MF, Razin S, Tully JG, Whitcomb RF. The Mycoplasmas, Volumes 1-4. Academic Press, N.Y., 1979.
8. http://www.bionique.com/ - Safe Cells Insights

Document#: DCF3013D
Edition#: 10
Effective Date: 07/15/2003
Title: **M-250 FINAL REPORT SHEET**

M-250 FINAL REPORT

Direct Specimen Culture
Procedure 3008, 3011, 3013

TO: WiCell QA
WiCell Research Institute

BTL SAMPLE ID#: **61671** P.O.#: DATE REC'D: **06/29/2010**

TEST/CONTROL ARTICLE:

WA09-WB0007 #5771

LOT#: **NA**

DIRECT CULTURE SET-UP (DAY 0) DATE: **06/30/2010**

INDICATOR CELL LINE (VERO) SEE DNA FLUOROCHROME RECORD SHEET

				DATE
THIOGLYCOLLATE BROTH	DAY 7	+	⊖	**07/07/2010**
	DAY 28	+	⊖	**07/28/2010**
BROTH-FORTIFIED COMMERCIAL				
0.5 mL SAMPLE	DAY 7	+	⊖	**07/07/2010**
6.0 mL BROTH	DAY 28	+	⊖	**07/28/2010**
BROTH-MODIFIED HAYFLICK				
0.5 mL SAMPLE	DAY 7	+	⊖	**07/07/2010**
6.0 mL BROTH	DAY 28	+	⊖	**07/28/2010**
BROTH-HEART INFUSION				
0.5 mL SAMPLE	DAY 7	+	⊖	**07/07/2010**
6.0 mL BROTH	DAY 28	+	⊖	**07/28/2010**

(See Reverse)

Document#: DCF3013D
Edition#: 10
Effective Date: 07/15/2003
Title: M-250 FINAL REPORT SHEET

SAMPLE ID#: **61671**		AEROBIC	MICROAEROPHILIC	DATE
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7	+ ⊖	+ ⊖	07/07/2010
	DAY 14	+ ⊖	+ ⊖	07/14/2010
	DAY 21	+ ⊖	+ ⊖	07/21/2010
AGAR PLATES-MODIFIED HAYFLICK	DAY 7	+ ⊖	+ ⊖	07/07/2010
	DAY 14	+ ⊖	+ ⊖	07/14/2010
	DAY 21	+ ⊖	+ ⊖	07/21/2010
AGAR PLATES-HEART INFUSION	DAY 7	+ ⊖	+ ⊖	07/07/2010
	DAY 14	+ ⊖	+ ⊖	07/14/2010
	DAY 21	+ ⊖	+ ⊖	07/21/2010
BROTH SUBCULTURES (DAY 7)		DATE: **07/07/2010**		
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7	+ ⊖	+ ⊖	07/14/2010
	DAY 14	+ ⊖	+ ⊖	07/21/2010
	DAY 21	+ ⊖	+ ⊖	07/28/2010
AGAR PLATES-MODIFIED HAYFLICK	DAY 7	+ ⊖	+ ⊖	07/14/2010
	DAY 14	+ ⊖	+ ⊖	07/21/2010
	DAY 21	+ ⊖	+ ⊖	07/28/2010
AGAR PLATES-HEART INFUSION	DAY 7	+ ⊖	+ ⊖	07/14/2010
	DAY 14	+ ⊖	+ ⊖	07/21/2010
	DAY 21	+ ⊖	+ ⊖	07/28/2010

RESULTS: No detectable mycoplasmal contamination

7/28/10
Date

Laboratory Director

M-250 Procedural Summary: The objective of this test is to ascertain whether or not detectable mycoplasmas are present in an in vitro cell culture sample, be it a primary culture, hybridoma, master seed stock or cell line. This procedure combines an indirect DNA staining approach to detect non-cultivable mycoplasmas with a direct culture methodology utilizing three different mycoplasmal media formulations. The indirect approach involves the inoculation of the sample into a mycoplasma-free VERO (ATCC) indicator cell line and performing a DNA fluorochrome assay after 72-120 hours of incubation. The direct culture aspect of the test utilizes three different mycoplasmal media including both broth and agar formulations. The sample is inoculated into each of the 3 broth formulations and also onto duplicate plates (0.1 mL/plate) for each of the 3 agar formulations. Subculture from broth to fresh agar plates is carried out after 7 days incubation. Agar plates are incubated aerobically and microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final report with signature of the Laboratory Director signifies that the required controls were performed concurrently with the test sample(s) as detailed in the referenced SOPs and that all test conditions have been found to meet the required acceptance criteria for a valid test, including the appropriate results for the positive and negative controls.

bionique Testing Laboratories

MYCOPLASMA TESTING SERVICES

Document ID #:	DCF3008A
Title:	**DNA FLUOROCHROME ASSAY RESULTS**
Effective Date:	3/24/10
Edition #:	07

DNA-FLUOROCHROME ASSAY RESULTS

Procedures 3008, 3009, 3011

Sample ID # **61671** **M-250** Date Rec'd: **06/29/2010** P.O. #

Indicator Cells Inoculated: Date/Initials: 7/1/10 / JA

Fixation: Date/Initials: 7/5/10 / HB

Staining: Date/Initials: 7/5/10 / HB

TEST/CONTROL ARTICLE:

WA09-WB0007 #5771

LOT# **NA**

WiCell QA
WiCell Research Institute

DNA FLUOROCHROME ASSAY RESULTS:

__X__ **NEGATIVE:** A reaction with staining limited to the nuclear region, which indicates no mycoplasmal contamination.

_____ **POSITIVE:** A significant amount of extranuclear staining which strongly suggests mycoplasmal contamination.

_____ **INCONCLUSIVE:**

_____ A significant amount of extranuclear staining consistent with low - level mycoplasmal contamination or nuclear degeneration.

_____ A significant amount of extranuclear staining consistent with bacterial, fungal or other microbial contaminant or viral CPE. Morphology not consistent for mycoplasmal contamination.

COMMENTS:

Date: 7/5/10 Results Read by: HB Date of Review: 7/6/10 Reviewed by: SAB

WiCell Cytogenetics Report: 001865-061810
WISC 5771

Report Date: June 28, 2010

Case Details:

Cell Line: *WA09-WB0007 (5771)*

Passage #: *22*

Date Completed: *6/28/2010*

Cell Line Gender: *Female*

Investigator:

Specimen: *hESC on Matrigel*

Date of Sample: *6/18/2010*

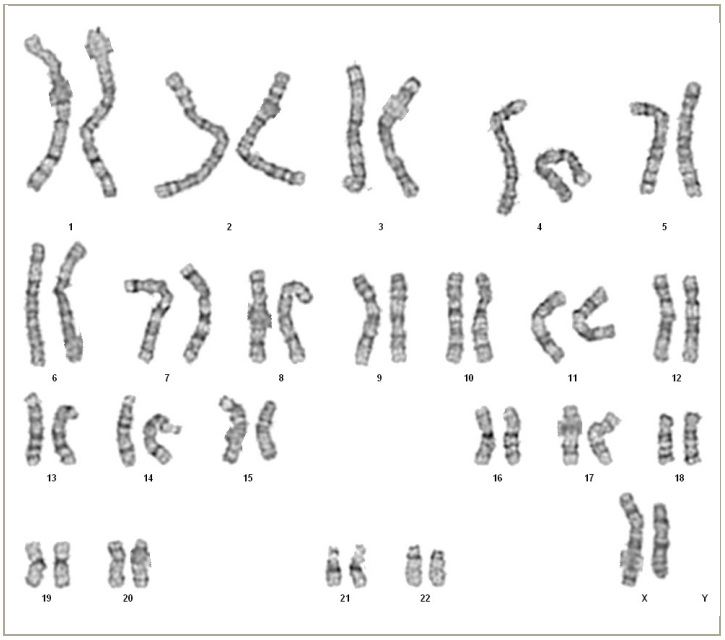
Tests,Reason for: *WB testing*

Results: *46,XX*

Completed by *MS, CG(ASCP), on 6/28/2010*

Reviewed and interpreted by *PhD, FACMG, on 6/28/2010*

Interpretation: *No clonal abnormalities were detected at the stated band level of resolution.*



Cell: *S01-04*

Slide: *B-20*

Slide Type: *Karyotyping*

# of Cells Counted: 20

# of Cells Karyotyped: 4

# of Cells Analyzed: 8

Band Level: 500-550

Results Transmitted by Fax / Email / Post **Date:**_____
Sent By:_____ **Sent To:**_____
QC Review By: _____ **Results Recorded:** _____