



Certificate of Analysis

Thaw and Culture Details

Cell Line Name	STAN241i-558C4	
WiCell Lot Number	WB68153	
Parent Material	STAN241i-558C4-DB38008	
Provider/Client	Stanford University – Laboratory of Dr. Thomas Quettermous	
Banked By	WiCell	
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 3 wells of a 6 well plate using mTeSR™ 1 and Cultrex®.	
Protocol	WiCell Feeder Independent Pluripotent Stem Cell Protocol	
Culture Platform Prior to Freeze	Medium: mTeSR™ 1	Matrix: Cultrex®
Passage Number	p17 Cells were cultured for 16 passages prior to freeze and post colony selection. Plated cells at thaw should be labeled passage 17.	
Date Vialied	22-June-2023	
Vial Label	STAN241i-558C4 p17 WB68153	
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.	



Certificate of Analysis

Results

Test Description	Test Provider	Test Method	Test Specification	Result
Karyotype	WiCell	G-T-L Banding performed on 20 metaphase cells	Expected karyotype	See Report
	Results: 46,XX Interpretation: This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.			
Post-Thaw Viable Cell Recovery	WiCell	Thaw using specified Thaw & Culture Recommendations	≥ 15 Undifferentiated Colonies prior to passage, ≤ 30% Differentiation prior to passage, and recoverable attachment after passage	Pass
Identity by STR	WiCell	PowerPlex 16 HS System by Promega™	Defines STR profile of deposited cell line	See Report
Mycoplasma	WiCell	PCR	Amplification of mycoplasma specific DNA detected with negative result	Pass
Sterility	Steris	Native Product Direct Transfer using FTM and TSB (ST/07)	Negative for growth following 14 days of culture	Pass

Testing Reported by Provider

The Provider stated that some or all of the additional analyses listed below may have been performed for this cell line. For more information, publication and dbGaP links, where available, are provided on the cell line specific web page on the WiCell website.

- RNA-Seq
- Whole Genome Sequencing

Approval Date	WiCell Quality Assurance Approval
30-November-2023	<div style="text-align: right; font-size: small;">11/30/2023</div> <div style="border: 1px solid black; padding: 2px; width: fit-content; margin: 0 auto;"> X HEB <small>HEB WiCell Quality Assurance Signed by: Bruner, Haley</small> </div>

Date Reported: Tuesday, October 31, 2023

Cell Line: STAN241i-558C4-WB68153

Submitted Passage #: 17

Date of Sample: 10/24/2023

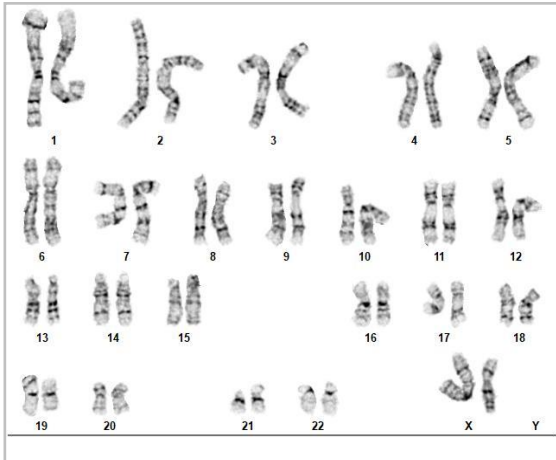
Specimen: Human iPSC

Results: 46,XX

Cell Line Sex: Female

Reason for Testing: LOT_RELEASE

Investigator: WiCell Stem Cell Bank, WiCell



Cell: 37

Slide: G01

Slide Type: Karyotype

Total Counted: 20

Total Analyzed: 8

Total Karyogrammed: 4

Band Resolution: 450 - 475

Interpretation:

This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.

Completed by: Jennifer Pecos, CG(ASCP)

Reviewed and Interpreted by: Vanessa Horner, PhD, FACMG

For internal use only

Date: _____ **Sent By:** _____ **Sent To:** _____ **QC Review By:** _____

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results of this assay are for research use only. Unless otherwise mutually agreed in writing, the services provided to you hereunder by WiCell Research Institute, Inc. ("WiCell") are governed solely by WiCell's Terms and Conditions of Service, found at www.wicell.org/privacyandterms. Any terms you may attach to a purchase order or other document that are inconsistent, add to, or conflict with WiCell's Terms and Conditions of Service are null and void and of no legal force or effect.



Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell
 Samples Received: 24Oct23, 26Oct23, 27Oct23
 STR Amplification Date: 07Nov23

Form SOP-89.01
 Version 11.0

Sample Name	UCSD012i-5-5-WB68191 p27	WA09-WB68167 p26	UCSD087i-6-4-WB68192 p20	WIC-WA09-MB-005 p28	STAN241i-558C4-WB68153 p17	STAN240i-558C3-WB68154 p17
WiCell CTR No. ¹	99362	99356	99355	99312	99311	99310
FGA	Identifying information has been redacted to protect donor confidentiality. If more information is required, please contact info@wicell.org					
TPOX						
D8S1179						
vWA						
Amelogenin						
Penta_D						
CSF1PO						
D16S539						
D7S820						
D13S317						
D5S818						
Penta_E						
D18S51						
D21S11						
TH01						
D3S1358						
Allelic Polymorphisms						
Matches*	72377	See Matches Comment	72176	See Matches Comment	99310	99311
Comments						

**Note: The STR profile of the following sample is a 100% match for the given sample/samples unless otherwise specified.*

¹ CTR No.: Characterization Test Request Number; also known as a laboratory accessioning number.



Short Tandem Repeat

Form SOP-89.01
Version 11.0

Requestor: WiCell Stem Cell Bank, WiCell
Samples Received: 24Oct23, 26Oct23, 27Oct23
STR Amplification Date: 07Nov23

Assay Description: STR analysis is performed using the PowerPlex 16 HS System by Promega™. Results are reported as 13 CODIS STR markers, Amelogenin for gender determination and two low-stutter, highly discriminating pentanucleotide STR markers.

Results: The genotypic profiles comprise a range of 24-29 allelic polymorphisms across the 15 STR loci analyzed.

Interpretation: The concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggests that the cells submitted correspond to the cell lines as named and were not contaminated with any other human cells or a significant amount of mouse feeder layer cells.

Sensitivity: Sensitivity limits for detection of STR polymorphisms unique to either this or other human cell lines is ~2-4%.

Matches: Samples 99356 and 99312 are a 100% match to each other and to 97827, 97437, 97371, 97171, 96184, 96183, 95823, 95822, 93654, 93595 and additional profiles. Additional matches can be provided upon request.

11/14/2023	11/14/2023	11/14/2023
<p>X Amber Kuhn</p> <hr/> <p>Tech #1 Characterization Signed by: Kuhn, Amber</p>	<p>X Anna Lisa Larson</p> <hr/> <p>Tech #2 Characterization Signed by: Larson, Anna Lisa</p>	<p>X Dawn Graham</p> <hr/> <p>QA Review Quality Assurance Signed by: Graham, Dawn</p>

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Mycoplasma Assay Report

PCR-based assay performed by WiCell
WiCell Stem Cell Bank, WiCell
01Nov23

Form SOP-83.01
Version 5.0

Sample Name	Result	Interpretation
STAN240i-558C3-WB68154 p17 (99310)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
STAN241i-558C4-WB68153 p17 (99311)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
WIC-WA09-MB-005 p28 (99312)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
UCSD087i-6-4-WB68192 p20 (99355)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
WA09-WB68167 p26 (99356)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
UCSD012i-5-5-WB68191 p27 (99362)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Positive (+) Control	Positive	
Negative (-) Control	Negative	

Assay Description
Sample is tested for presence of mycoplasma using EZ-PCR™ Mycoplasma Detection Kit (Sartorius).

11/2/2023	11/2/2023	11/2/2023
X John Raff	X Kaylie Petersen	X Dawn Graham
Tech #1 Characterization Signed by: Raff, John	Tech #2 Characterization Signed by: Petersen, Kaylie	QA Review Quality Assurance Signed by: Graham, Dawn

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A gel image is available upon request.

Native Product Sterility Report



WiCell
504 S Rosa Road, Rm 101
Madison, WI 53719

SAMPLE #: 23070508
DATE RECEIVED: 13-Jul-23
TEST INITIATED: 17-Jul-23
TEST COMPLETED: 31-Jul-23

SAMPLE NAME / DESCRIPTION: JHU189i-DB41401
JHU192i-DB36782
JHU215i-DB36857
JHU227i-DB37013
JHU230i-DB37025
JHU081i-DB41140
JHU150i-DB41359
STAN240i-558C3-WB68154
STAN241i-558C4-WB68153
H1 OCT4-EGFP-2A-C.3-WB68155
H1 MYH11-NLuc-tdTomato-WB68133
UCSD247i-LQT1-2-WB68158

UNIQUE IDENTIFIER: N/A

TEST RESULTS:

# Tested	# Positives (Growth)	- Control
12	0	2 Negatives

TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
12	TSB	40	20-25	14
12	FTG	40	30-35	14

REFERENCE: Processed according to LAB-003: Sterility Test Procedure

PD #: 000053

TEST METHODOLOGY: USP - Direct Transfer

COMMENTS: NA

AUTHORIZED BY 

DATE 02 Aug 2023

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests. Results applied to samples as received.