

Thaw and Culture Details

Cell Line Name	WA09
WiCell Lot Number	WIC-WA09-RB-002
Parent Material	WIC-WA09-MB-001
Provider	WiCell
Banked By	WiCell
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 3 wells of a 6 well plate.
Culture Platform	Feeder Independent
	Medium: mTeSR™1
	Matrix: Matrigel®
Protocol	WiCell Feeder Independent mTeSR™1 Protocol
Passage Number	p29 These cells were cultured for 28 passages prior to freeze, 6 of them in mTeSR1/Matrigel. WiCell adds +1 to the passage number to best represent the overall passage number of the cells at thaw.
Date Vialed	02-September-2009
Vial Label	WIC WA09 RB 002 P29 MW 02 SEPT 2009 SOP-CC-038A
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

Testing Performed by WiCell

Test Description	Test Provider	Test Method	Test Specification	Result
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	≥ 15 Undifferentiated Colonies, ≤ 30% Differentiation	Pass
Identity by STR	UW Molecular Diagnostics Laboratory	PowerPlex 1.2 System by Promega	Consistent with known profile	Pass
Sterility - Direct transfer method	Apptec	30744	Negative	Pass
Mycoplasma	Bionique	M250	No contamination detected	Pass
Karyotype by G-banding	WiCell	SOP-CH-003	Normal karyotype	Pass
Flow Cytometry for ESC Marker Expression	UW Flow Cytometry Laboratory	SOP-CH-101 SOP-CH-102 SOP-CH-103 SOP-CH-105	Report - no specification	See report



Approval Date	Quality Assurance Approval
31-January-2011	10/4/2016 X AMK AMK Quality Assurance Signed by Klade, Anjelica





Short Tandem Repeat Analysis*

Sample Report: 6520-STR

UW HLA#: 62095

Sample Date: 11/20/09

Received Date: 11/20/09

Requestor: WiCell Research Institute

Test Date: 11/23/09

File Name: 091124

Report Date: 11/27/09

Sample Name: (label on tube) 6520-STR

Description: DNA Extracted by WiCell

255.77 ug/mL; 260/280 = 1.90

Locus	Repeat #	STR Genotype
D16S539	5, 8-15	12,13
D7S820	6-14	9,11
D13S317	7-15	9,9
D5S818	7-15	11,12
CSF1PO	6-15	11,11
TPOX	6-13	10,11
Amelogenin	NA	X,X
TH01	5-11	9.3,9.3
vWA	11, 13-21	17,17

Comments: Based on the DNA 6520-STR dated and received on 11/20/09 from WI Cell, this sample (UW HLA# 62095) matches exactly the STR profile of the human stem cell line H9 comprising 12 allelic polymorphisms across the 8 STR loci analyzed. No STR polymorphisms other than those corresponding to the human H9 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggest that the 6520-STR DNA sample submitted corresponds to the H9 stem cell line and it was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells. Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is $\sim 5\%$.

anager Date

HLA/Molecular Diagnostics Laboratory

HLA/Molecular Diagnostics Laboratory

* Testing to assess engraftment following bone marrow transplantation was accomplished by analysis of human genetic polymorphisms at STR loci. This methodology has not yet been approved by the FDA and is for investigational use only.

File: Final STR Report

Test Facility:

This report is confidential. No part may be used for advertising or public announcement without written permission. Results apply only to the sample(s) tested.



Report Number 819787 Page 1 of 1

October 16, 2009 P.O. #:

WiCell Research Institute

STERILITY TEST REPORT

Sample Information:

hES Cells

WIC-WA09-RB-002 # 3503

Date Received: Date in Test:

September 24, 2009 October 01, 2009

Date Completed:

October 15, 2009

Test Information:

Test Codes: 30744, 30744A Immersion, USP / 21 CFR 610.12 Procedure #: BS210WCR.201

TEST PARAMETERS	PROI	PUCT	
Approximate Volume Tested	0.5 mL	0.5 mL	
Number Tested	2	2	
Type of Media	SCD	FTM	
Media Volume	400 mL	400 mL	
Incubation Period	14 Days	14 Days	
Incubation Temperature	20 °C to 25 °C	30 °C to 35 °C	
RESULTS	2 NEGATIVE	2 NEGATIVE	

QA Reviewer

Téchnical Reviewer

Date

Testing conducted in accordance with current Good Manufacturing Practices.





BIONIQUE® TESTING LABORATORIES, INC.

APPENDIX

BIONIQUE® TESTING LABORATORIES, INC.

Document ID #: DCF9002E

Title: QUALITY ASSURANCE REPORT - GMP

Effective Date: 05/21/09 Edition #: 01

QUALITY ASSURANCE REPORT - GMP

TEST PERFORMED	PROCEDURAL REFERENCE		TEST PERFORMED	PROCEDURAL REFERENCE
M-250M-300M-350	SOP's 3008, 3011, 3013 SOP's 3008, 3014 SOP's 3008, 3014, 3015	1 -	☐ M-700 ☐ M-800	SOP's 3008, 3009, 3010 SOP's 3008, 3011, 3016
Bionique Sample ID	#(s) <u>59388</u>		How were employed	

This testing procedure was performed in compliance with the FDA's Current Good Manufacturing Practice (cGMP) standards (to the extent that the regulations pertain to the procedures performed) as specified in the Code of Federal Regulations, Title 21 Parts 210 and 211 [21 CFR 210 & 211]. All related records derived from the test procedures have been reviewed by the Quality Assurance Department. The individual's signature below verifies that the methods and procedures referenced above have been followed and that the Final Report accurately reflects the raw data generated during the course of the procedures. All records, including raw data and final reports are archived on site for a minimum of seven years.

The specified test's procedures determine the intervals at which samples are inspected. The medium used for testing must pass quality control mycoplasmal growth promotion testing and sterility testing. Traceability of all of the components used is assured and supporting documentation can be supplied upon request.

Quality Assurance Review Date	e: 12/10/0 ^C		
Reviewed By	(QA Assistant:		

NOTE:

- 1. Prior to receipt at Bionique[®] Testing Laboratories, Inc., the stability of the test article is the responsibility of the company submitting the sample. Bionique Testing Laboratories Inc. will assume responsibility for sample stability following receipt and prior to being placed on test.
- 2. This test is for the detection of microbiological growth and does not require statistical validation.

APPENDIX

BIONIQUE® TESTING LABORATORIES, INC.

Document ID #: DCF9002E

Title: QUALITY ASSURANCE REPORT - GMP

Effective Date: 05/21/09

Edition #: 01

REFERENCES

Regulatory:

- 1. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Code of Federal Regulations [CFR], Title 21 CFR Part 210, Current Good Manufacturing Practice in Manufacturing, Processing, Packing, or Holding of Drugs; General. FDA. Office of the Federal Register, National Archives and Records Department.
- 2. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Code of Federal Regulations [CFR], Title 21 CFR Part 211, Current Good Manufacturing Practice for Finished Pharmaceuticals. FDA. Office of the Federal Register, National Archives and Records Department.
- 3. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Points to Consider in the Characterization of Cell Lines Used to Produce Biologicals, Director, Center for Biologics Evaluation and Research, FDA. May, 1993. Docket No. 84N-0154.
- 4. Department of Health and Human Services, Food and Drug Administration (USA) [FDA]. Code of Federal Regulations [CFR], Title 21 CFR Part 610.30, General Biological Products Standards; Subpart D, Test for Mycoplasma. FDA. Office of the Federal Register, National Archives and Records Department.

General:

- 1. Barile MF, Kern J. Isolation of Mycoplasma arginini from commercial bovine sera and its implication in contaminated cell cultures. Proceedings of the Society for Experimental Biology and Medicine, Volume 138, Number 2, November 1971.
- 2. Chen, T.R. In situ detection of mycoplasma contamination in cell cultures by fluorescent Hoechst 33258 stain. Experimental Cell Research, 104: 255-262, 1977.
- 3. Carolyn K. Lincoln and Daniel J. Lundin. Mycoplasma Detection and Control. U. S. Fed. for Culture Collections Newsletter, Vol. 20, Number 4, 1990.
- 4. Fetal Bovine Serum; Proposed Guideline. National Committee For Clinical Laboratory Standards (NCCLS), Vol. 10, Number 6, 1990. (NCCLS publication M25-P).
- 5. McGarrity GJ, Sarama J, Vanaman V. Cell Culture Techniques. ASM News, Vol. 51, No. 4, 1985.
- 6. Tully JG, Razin S. Methods in Mycoplasmology, Volumes I and II. Academic Press, N.Y., 1983.
- 7. Barile MF, Razin S, Tully JG, Whitcomb RF. The Mycoplasmas, Volumes 1-4. Academic Press, N.Y., 1979.
- 8. http://www.bionique.com/ Safe Cells Insights



BIONIOUE TESTING LABORATORIES, INC.

APPENDIX IV

Page 1 of 2

Document#: Edition#:

DCF3013D

10

Effective Date:

07/15/2003

Title:

M-250 FINAL REPORT SHEET

M-250 FINAL REPORT

Direct Specimen Culture Procedure 3008, 3011, 3013

TO: Wicell QA WiCell Research Institute

BTL SAMPLE ID#: 59388

P.O.#:

DATE REC'D:

11/12/2009

TEST/CONTROL ARTICLE:

WIC-WA09-RB-002-D#6520

LOT#:

NA

(See Reverse)

DIRECT CULTURE SET-UP (DAY 0)	DATE	11/12/2009	
INDICATOR CELL LINE (VERO)	SEE DNA FLUOROCH	ROME RECORD SHEET	
		DATE	
THIOGLYCOLLATE BROTH	DAY 7 +		09
*	DAY 28 +		09
BROTH-FORTIFIED COMMERCIAL			
0.5 ml SAMPLE	DAY 7 +	○ <u>11/19/20</u>	09
6.0 mL BROTH	DAY 28 +	<u>12/10/20</u>	09
BROTH-MODIFIED HAYFLICK			
0.5 mL SAMPLE	DAY 7 +	<u>11/19/20</u>	09
6.0 mL BROTH	DAY 28 +		09
BROTH-HEART INFUSION			v
0.5 mL SAMPLE	DAY 7 +		09
6.0 mL BROTH	DAY 28 +	<u>12/10/20</u>	09

Document#:

DCF3013D

Edition#:

10

Effective Date:

07/15/2003

Title:

M-250 FINAL REPORT SHEET

SAMPLE ID#: 59388	F	AEROBIC MICR	OAEROPHILIC	DATE
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7 DAY 14 DAY 21	+ ⁽⁻⁾ + ⁽⁻⁾ + ⁽⁻⁾	+ (5)	$\frac{11/19/2009}{11/26/2009}$ $\frac{12/03/2009}{12}$
AGAR PLATES-MODIFIED HAYFLICK	DAY 7 DAY 14 DAY 21	+ (D) + (D) + (C)	+ (-) + (-) + (-)	$\frac{11/19/2009}{11/26/2009}$ $\frac{12/03/2009}{12}$
AGAR PLATES-HEART INFUSION	DAY 7 DAY 14 DAY 21	+ © + © + ©	+ (D) + (O) + (O)	$\frac{11/19/2009}{11/26/2009}$ $\frac{12/03/2009}{12}$
BROTH SUBCULTURES (DAY 7)	D	ATE: <u>11/19/2</u>	009	
AGAR PLATES-FORTIFIED COMMERCIAL	DAY 7 DAY 14 DAY 21	+ (D) + (D) + (D)	+ (<u>)</u> + (<u>)</u> + (<u>)</u>	11/26/2009 12/03/2009 12/10/2009
AGAR PLATES-MODIFIED HAYFLICK	DAY 7	+ 😊	+ 🐑	11/26/2009
	DAY 14 DAY 21	+ 🕥	+ 🕞	12/03/2009

RESULTS: No detectable mycoplasmal contamination

<u> 14/0/09</u> Date

Laboratory Director

Ph.D.

M-250 Procedural Summary: The objective of this test is to ascertain whether or not detectable mycoplasmas are present in an in vitro cell culture sample, be it a primary culture, hybridoma, master seed stock or cell line. This procedure combines an indirect DNA staining approach to detect non-cultivable mycoplasmas with a direct culture methodology utilizing three different mycoplasmal media formulations. The indirect approach involves the inoculation of the sample into a mycoplasma-free VERO (ATCC) indicator cell line and performing a DNA fluorochrome assay after 72-120 hours of incubation. The direct culture aspect of the test utilizes three different mycoplasmal media including both broth and agar formulations. The sample is inoculated into each of the 3 broth formulations and also onto duplicate plates (0.1 mL/plate) for each of the 3 agar formulations. Subculture from broth to fresh agar plates is carried out after 7 days incubation. Agar plates are incubated aerobically and microaerophillically in order to detect any colony forming units morphologically indicative of mycoplasmal contamination. Issuance of the final report with signature of the Laboratory Director signifies that the required controls were performed concurrently with the test sample(s) as detailed in the referenced SOPs and that all test conditions have been found to meet the required acceptance criteria for a valid test, including the appropriate results for the positive and negative controls.



BIONIQUE TESTING LABORATORIES, INC

Edition #: Effective date: Title:	DCF3008A 06 9/17/2003 DNA FLU	OROCHROME	ASSA	Y RESUI	LTS		
ν 1 - 8 γ	DNA-FI	UOROCHROME A					
Sample ID # <u>59388</u>	<u>M-250</u>	Date Rec'd:	11/12	2/2009	P.O. #		
Indicator Cells Inoculated:	Date/Initials:	11/13/09	/	tts	- 20		
Fixation:	Date/Initials:	11/17/09	/	K6			
Staining:	Date/Initials:	11/17/09	/	K6			
TEST/CONTROL ARTICLE:				8			
WIC-WA09-RB-002-I	0#6520						
LOT# <u>NA</u>							
				Phone: Fax #:		5	
DNA FLUOROCHROM	E ASSAY RES	ULTS:					
DNA FLUOROCHROM:	: A reaction	ULTS: n with staining lasmal contami		Fax #:	uclear reg	ion, whic	h indi
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POSITIVE:	A reaction no mycop A signific mycoplas JSIVE: A signific mycoplas A signific fungal or	n with staining clasmal contaminate amount of emal contaminate ant amount of emal contaminate ant amount of emal contaminate ant amount of emal contaminate and amount of emal contaminate	extranution. extranution or extranucli cont	Fax #: d to the man. clear stain nuclear stain aminant	ining which ining considegenerations consions or viral CF	ch strongl stent with ion. istent wit	ly sug h low h bac



WiCell Cytogenetics Report: 001434-102809 WISC 6520

Report Date: November 06, 2009

Case Details:

Cell Line: WIC-WA09 (6520)

Passage #: 30

Date Completed: 11/6/2009
Cell Line Gender: Female

Investigator: cGMP

Specimen: hESC on Matrigel Date of Sample: 10/28/2009

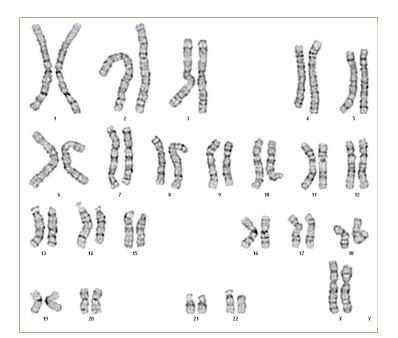
Tests, Reason for: Research Bank Testing for cGMP

Results: 46.XX

Completed by CLSp(CG), on 11/5/2009

Reviewed and interpreted by PhD, FACMG, on 11/6/2009

Interpretation: No clonal abnormalities were detected at the stated band level of resolution.



Cell: S01-01

Slide: A-5

Slide Type: Karyotyping

of Cells Counted: 40

of Cells Karyotyped: 4

of Cells Analyzed: 8

Band Level: 475-525

Results Transmitted by Fax / Email / Post Sent By: QC Review By: Date:_

Sent To:

Results Recorded:



0.55

0.00

antigen2: SSEA3 TRA1-60 TRA1-81

Oct-4

SSEA1

Procedures performed: SOP-CH-101 SOP-CH-102 SOP-CH-103 SOP-CH-105

96.10

0.15

Cell Line: WIC-WA09-RB-002

99.20

99.85

Passage 30

0.27

0.12

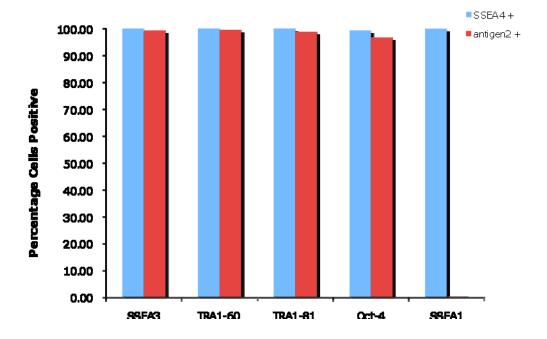
Sample ID: 6717-FAC

Date of: (mm/dd/yy) acquisition: 12/14/10 file creation: 12/23/10 file submission: 12/23/10

96.65

0.15

SSEA4 -	SSEA4 +	SSEA4 +	SSEA4 -	ALL	ALL
antigen2 +	antigen2 +	antigen2 -	antigen2 -	SSEA4 +	<u>antigen2 +</u>
0.04	99.20	0.74	0.00	99.94	99.24
0.07	99.40	0.50	0.02	99.90	99.47
0.06	98.70	1.22	0.02	99.92	98.76



3.10

99.70

